

Spatial analysis of epidemics: Disease gradients and patterns

Space....The Frontier....Finally.

- Epidemics are dynamic population processes in time *and space*
- We have spent a few weeks dealing exclusively with temporal dynamics
- However, diseases usually spread in space as they increase in time
- Because of spread, disease intensity is not the same everywhere in a field or plot -- there is a *pattern* to disease
- Spread *may or may not* be relatively easy to measure and quantify.

"Given that the transmission of pathogens leading to disease requires the close juxtaposition of a susceptible [healthy, disease-free] individual with an infected conspecific, vector, or environmental source of pathogens, transmission dynamics are inherently spatial processes."

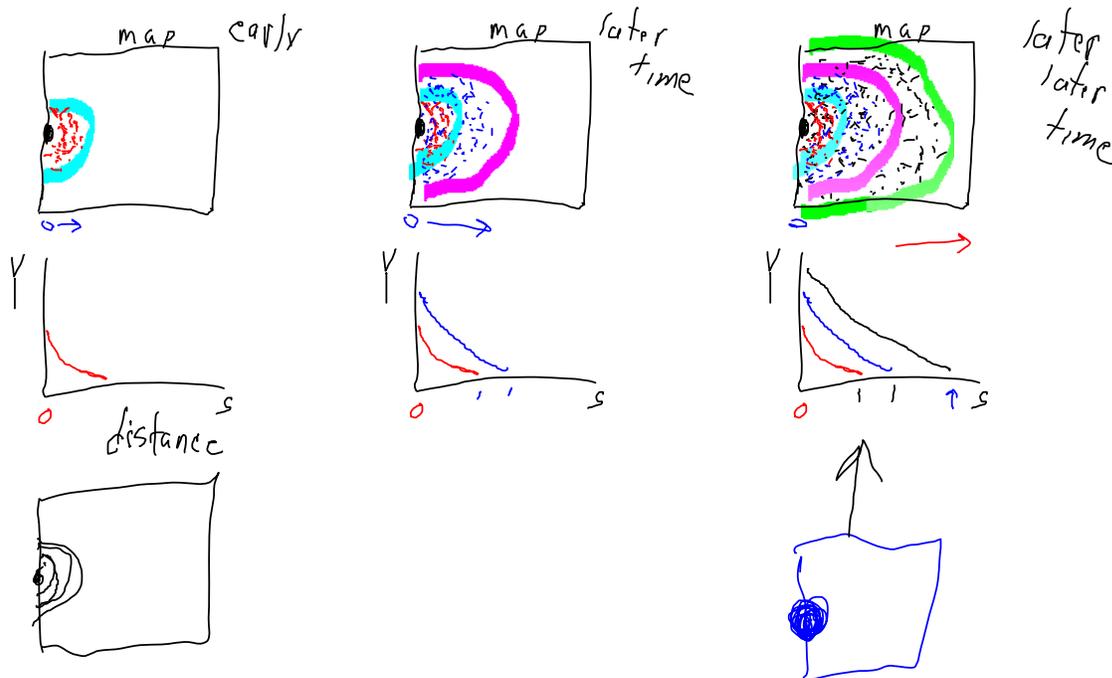
-- Ostfeld, Glass, and Keesing (2005). Spatial epidemiology: an emerging (or re-emerging) discipline. *Trends in Ecology and Evolution* 20: 328-336.

From HLIR model (coupled differential equations):

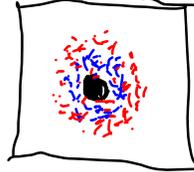
$$dH/dt = -\beta IH = -(\alpha\theta\psi)IH \quad \text{or} \quad dY/dt = +\beta IH = +(\alpha\theta\psi)IH$$

θ : Ultimately depends on distance between I and H

Results of spatial analysis allows, ultimately, for the expansion of temporal models to spatio-temporal models of epidemics (later)

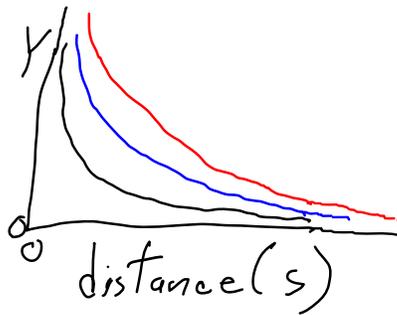


Some scenarios



spread

time
1
2
3



see figure 7.1, 7.17



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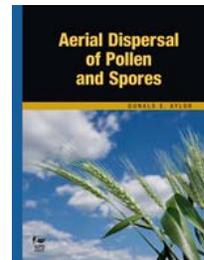
Concepts

- Disease spread implies movement, but diseased plants (leaves, etc.) do not move (typically), just the inoculum
- We consider spread to be the result of dispersal
- **Dispersal:**
 - Movement of 'inoculum' (such as spores, infectious units, propagules, viruliferous vectors) from one place to another
 - Movement of units of inoculum from the place they were formed to other locations
 - Involves **liberation, transport, deposition**
 - Pioneer: **P. H. Gregory** (see his pioneering book)
 - See articles by **Don Aylor**, A. McCartney, L. Huber, S. Isard, others
- **Because of dispersal, the intensity of disease varies systematically with distance from the source of inoculum**
 - There is more disease in some places than others
- We primarily deal with disease intensity and not the 'inoculum'
 - There is a vast literature on the physics of inoculum dispersal (a **very worthy topic**), but we do not discuss here (an entire course can be devoted to this topic!)



removal slight landing → aerobiology

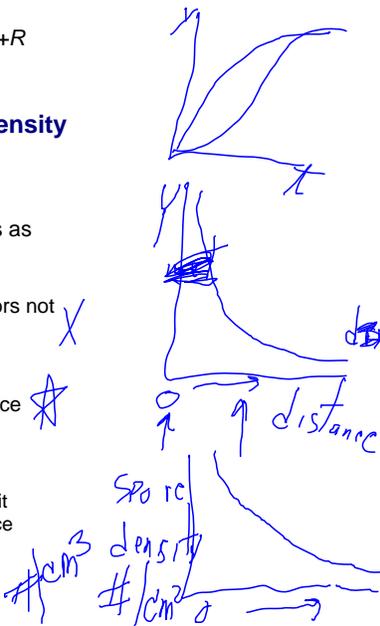
Don Aylor (2017). APS Press.



versus distance

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- For temporal dynamics, we always considered the **disease progress curve** (y versus t , or Y versus t , or L , I , R , and $L+I+R$ versus t)
- For spatial dynamics, we consider the **disease gradient**:
The change (generally, decline) in disease intensity over distance from an inoculum source
Intensity: incidence, severity (including counts)
- **Disease gradients** can have multiple causes (which serves as one convenient way of classifying them)
 - **Environmental gradient**
Disease gradient due to physical or other external factors not related to dispersal (or the biology of the disease)
 - **Dispersal gradient**
Disease gradient caused by variation in deposition of 'inoculum' in relation to distance from an inoculum source
The type of gradient of interest to us
 - **Deposition gradient**
 - (not a disease gradient at all). 'Inoculum' deposited per unit area (or length) in relation to distance from inoculum source
 - ◻ Deposition gradients *lead* to dispersal gradients

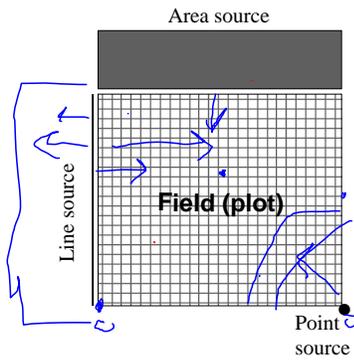


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- **Disease gradients due to dispersal (i.e., dispersal gradients) are of importance here**
- More concepts: **inoculum source**
 - For polycyclic diseases, any infected (specifically, infectious [I]) individual is an inoculum source for other disease-free (H) individuals
 - For monocyclic diseases, inoculum source can be infected individuals at another location ('external' to the current epidemic) or in another year; or spores in the soil, or spores originating elsewhere (another epidemic)
 - But, to *characterize* (i.e., study) dispersal gradients, we need to be more restrictive in what we consider an **inoculum source (an operational definition)**:
 - **A spatially-restricted concentration of spores, infectious units, other units of 'inoculum', or diseased individuals that can produce 'inoculum' (disease focus)**
 - ◻ Generally, the source is much smaller in area than the area of interest (field) (in our operational definition)
 - ◻ Typically consider the source as 'separate' from the area of interest (for convenience of experimentation only)
 - Classify inoculum sources based on size and geometry



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Kinds of inoculum sources

(see Chapter 7 in MHV for more detailed definitions)

You must know definitions of these terms .

Point source: width (size) of source is $< 1\%$ of the distance over which spread is measured

Line source: point source stretched over the entire width (length) of the field or plot, perpendicular to the direction of spread

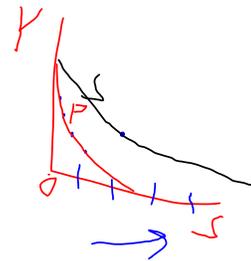
Area source: A "line source with depth"; width is considerably more than 1% of the distance of the field.

← Partial definitions

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More classifications of dispersal gradients (or disease gradients resulting from dispersal gradients):

- **Primary gradient:**
 - All diseased individuals (infections) are due to spores (or other infectious units) originating at the original inoculum source
 - Essentially: dealing with **primary infections** (whether the epidemic is monocyclic or polycyclic)
 - May continue over short or long times
- **Secondary gradient:**
 - Diseased individuals (infections) are due to inoculum produced outside of the original inoculum source (and in the host population of interest)
 - Essentially: dealing with **secondary infections** and polycyclic epidemics
- Some gradients may be a mixture of both



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Models of disease (dispersal) gradients

- As with disease progress curves, epidemiologists routinely use models (both simple and very complex) to characterize disease gradients
- Most common (and simple) models with acceptable biological realism are deterministic, either in the form of differential equations, integrated nonlinear models, or linearized equations
- Notation:**
 - s**: distance (as in space)
 - Y**: disease intensity in absolute units (lesions, infections), or even spore density
 - Since many gradients are concerned with numbers of lesions (or even spores), we start with Y
 - Y(s)**: Y at distance **s**; also Y_s
 - y**: disease intensity as a proportion $Y/M = y$
 - a** and **b** (with subscripts): parameters
 - dY/ds**: absolute rate of change in Y with change in distance (s); steepness of the gradient
- Model forms will be (should be!) quite familiar to you (by now)
 - These do not necessarily align with monocyclic and polycyclic diseases (so, be careful in interpretation!)

$$\frac{dY}{ds}$$

$$\frac{dY}{ds}$$

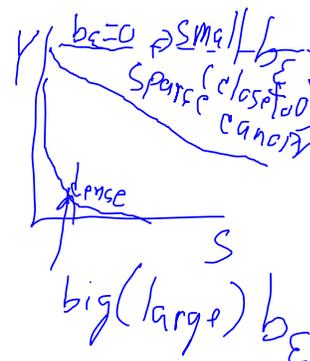
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Exponential model (negative exponential)

→ Gregory model

- First used by Frampton (1942) and Gregory and Read (1949) for gradients
 - But, many call it the Kiyosawa & Shiyomi model (after their 1972 paper)
- $\frac{dY}{ds} = -b_E Y$
 - Absolute rate is *negative*, because Y declines with distance
 - Rate is proportional to Y, meaning: the higher the Y, the steeper the decline in Y (*the greater the density of disease, the greater the decline over distance*)
 - b_E**: **dispersal or spread parameter**; units of 1/distance
 - Function of dispersal process (physics) and properties of host and pathogen
 - Higher susceptibility, or higher aggressiveness (higher infection efficiency), or less dense crop, for example, may result in lower b_E (closer to 0)
 - Large b_E means steep gradient

$$\frac{dY}{ds} = -b_E Y$$



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Exponential model

- $dY/ds = -b_E Y \rightarrow Y = a_E \exp[-b_E s]$
 - a_E :
 - parameter; constant of integration; value of Y at $s=0$, $Y(0)$ or Y_0 (because $\exp[-b_E 0] = \exp[0] = 1$)
 - Indicator of height of the curve
 - Measure of **'source strength'** -- *estimated* (predicted) amount of inoculum or disease at the source
 - Remember: in experiments, one often does not directly observe Y at the source

$$Y = a_E e^{-b_E s}$$

$$\begin{aligned} a_E e^{-b_E \cdot 0} \\ &= a_E (e^0) \\ &= a_E \cdot 1 = a_E \end{aligned}$$

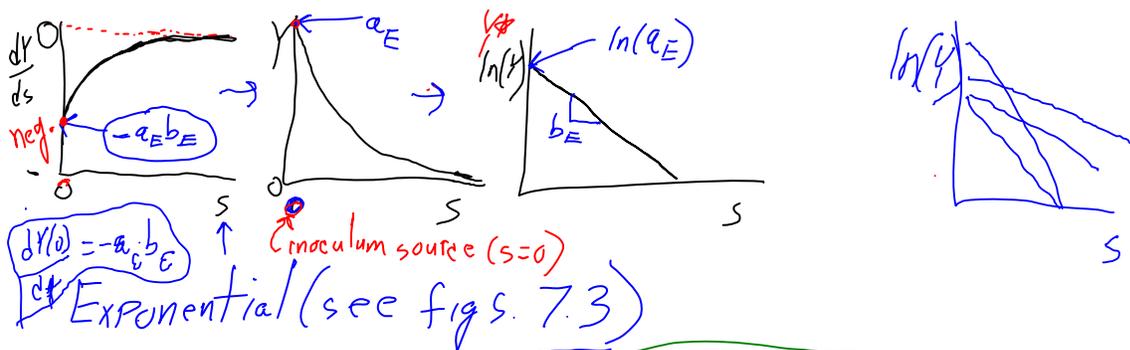
- $\ln(Y) = \ln(a_E) - b_E s$

- $\ln(a_E)$ $\ln(a_E)$ $\ln(a_E)$

- Linear model in terms of $\ln(a_E)$ and b_E : $Y^* = a^* - b^* s$

- Note: in regression analysis: it is assumed that there are pluses (+) between terms. Thus, one will obtain a negative slope, even though b is really positive (the minus is part of the model)

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$$\begin{aligned} \frac{dY}{ds} &= -b_E Y \rightarrow Y = a_E e^{-b_E s} \\ \ln(Y) &= \ln(a_E) - b_E s \\ Y^* &= a^* - b^* s \end{aligned}$$

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Power model (power-law or inverse power model)

Compare with
(negative)
exponential model for
disease gradient,

- Sometimes called the **Gregory** model because he was an early user of the equation (reviewed in 1968)

- $dY/ds = -b_p Y/s$

- Absolute rate is negative, because Y declines with distance
- Rate is proportional to Y, meaning: the higher the Y, the steeper the decline
- Rate is also inversely proportional to s, meaning: rate gets smaller (closer to 0) at increasing distance (see the 'dilution' argument later)
 - (but Y also gets smaller; thus more complicated)
- b_p : dispersal or spread parameter; unitless (1/distance is explicitly in the model)
 - Function of dispersal process and properties of host and pathogen
 - Direct interpretation is trickier (as will be shown later)

$$\frac{-b_p Y}{s}$$

$$s; \quad \frac{1}{1} \quad \frac{1}{10} \quad \frac{1}{100}$$

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Power model (power-law or inverse power model)

- $dY/ds = -b_p Y/s$

$$\longrightarrow Y = a_p s^{-b_p}$$

$$a_p e^{-b_p s}$$

Exp. model (compare)

- a_p :

- A 'source-strength'-type parameter
- Indicator of height of the curve
- Estimated (predicted) **value of Y at s = 1** (not at s=0): $Y(1)$ or Y_1
 - Insert 1 for s in equation, and one gets $a_p \cdot 1 = a_p$
 - If s is in meters, a_p is Y at 1 m; if s is in kilometers, it's Y at 1000 m!
 - Use small distance scale for Y near the source (represent s in centimeters)
- At s = 0, model is undefined (can't divide by 0; can't raise 0 to a negative value)
 - Y approaches $+\infty$ in the limit as s approaches 0
 - Thus, **Y at the source is ∞** (impossible)
 - But model may be excellent at $s > 0$

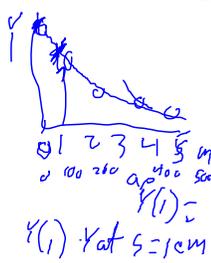
- $\ln(Y) = \ln(a_p) - b_p \ln(s)$

Linear model: $Y^* = a^* - b \cdot s^*$

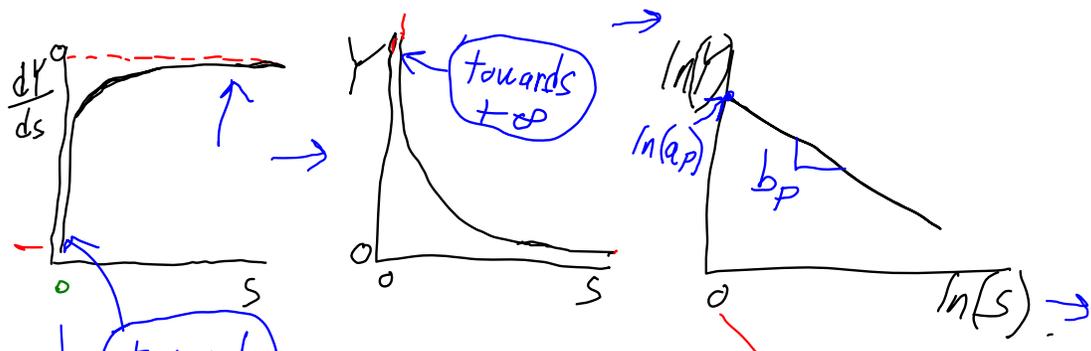


$$a_p 0^{-b_p} = \infty$$

$$a_p \cdot 1^{-b_p} = a_p$$



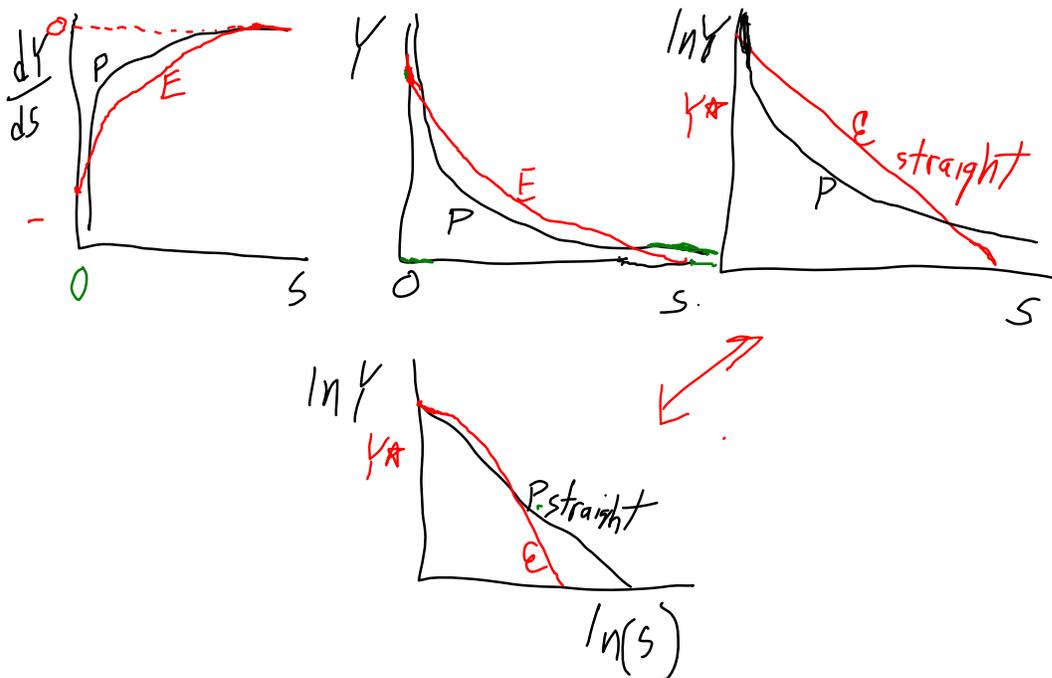
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Power
see fig. 7.4

$\ln(s) = 0$
 $s = 1$
 $\ln(r) = 0$

$\frac{dY}{ds}$	$s=1$
	0.1
	0.01
	0.001
	0.0001
	0.00001
R	\downarrow

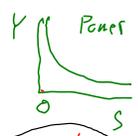


Modified power model (add a constant, λ)

- $dY/ds = -b_p Y/(s+\lambda) \rightarrow Y = a_p (s+\lambda)^{-b_p}$
- $\ln(Y) = \ln(a_p) - b_p \ln(s+\lambda)$
- a_p : Indicator of height of the curve
- λ : either arbitrary constant or approximate size of the source
- At $s=0$, all relevant calculations are possible...

$a_p s^{-b_p}$

$\frac{dY}{ds} = \frac{-b_p Y}{s+\lambda}$



$Y_0 = a_p (\lambda)^{-b_p}$

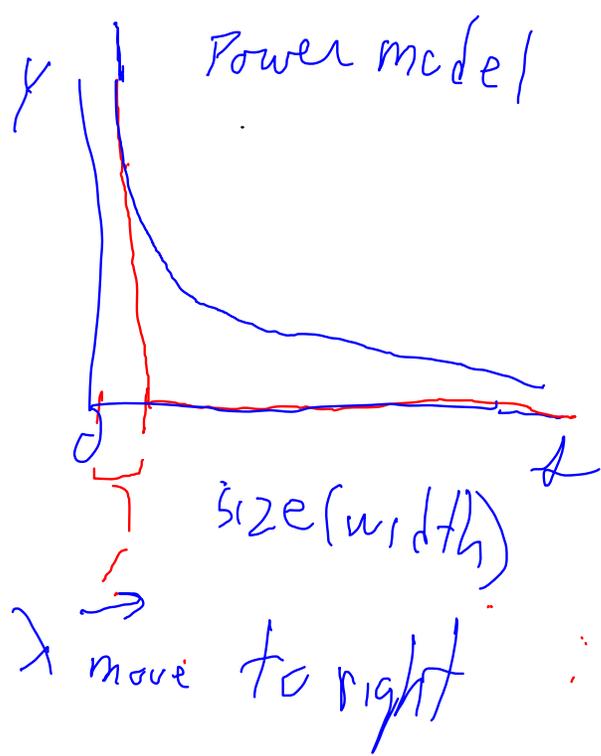
is the 'true' source strength (comparable to a_E of exponential)

$Y = a_p (s+\lambda)^{-b_p}$
 $a_p \cdot 1^{-b_p} = a_p$

Exp.
 source strength
 $= a_E$

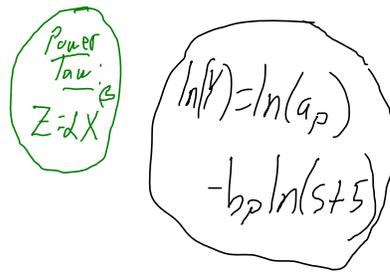
- If one inserts $s = 1-\lambda$ into model, one gets a_p
 - Thus, a_p is predicted Y at $1-\lambda$ units of distance from center of source (shifting disease gradients λ to the left $(-\lambda)$).
 - Think, then, of λ as 'diameter' of inoculum source

$q_p(0+\lambda)^{-b_p} = q_p(\lambda)^{-b_p}$



Modified power model (add a constant, λ)

- $dY/ds = -b_p Y / (s + \lambda)$ $Y = a_p (s + \lambda)^{-b_p}$
- $\ln(Y) = \ln(a_p) - b_p \ln(s + \lambda)$ *nonlinear*
- a_p : Indicator of height of the curve
- λ : either arbitrary constant or approximate size of the source
- Plots are similar to standard power model (except very close to source)
- Standard power model is just a special case (with $\lambda = 0$)
- Ad hoc method when one needs a finite source-strength estimate:
- For λ : use one-half the distance between 0 and the first distance with observed Y



Or, fit model with non-linear regression (model is NOT linear because λ is inside the log function)

$\lambda = 20 \rightarrow 9.99999$

$\ln(Y) = \ln(a_p) - b \ln(s + \lambda)$

$\ln(s + 1.0) \leftarrow$ choose λ :

$\ln(s + 1) \leftarrow$ highest R^2

$\ln(s + 2)$

\downarrow

$\ln(s + 9)$

$\ln(s + 9.9)$

lowest MSE (SSE)

Thus, there are two **fundamental** model forms for disease gradients

Exponential:

$$dY/ds = -b_E Y$$

$$Y = a_E e^{-b_E s}$$

$$\ln(Y) = \ln(a_E) - b_E s$$

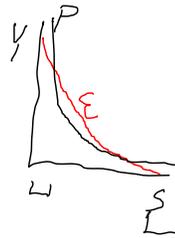
Modified power (λ), or power:

$$dY/ds = -b_P Y / (s + \lambda)$$

$$Y = a_P (s + \lambda)^{-b_P}$$

$$\ln(Y) = \ln(a_P) - b_P \ln(s + \lambda)$$

$\lambda = 0$
regular power



Spore **deposition**
(low air turbulence)

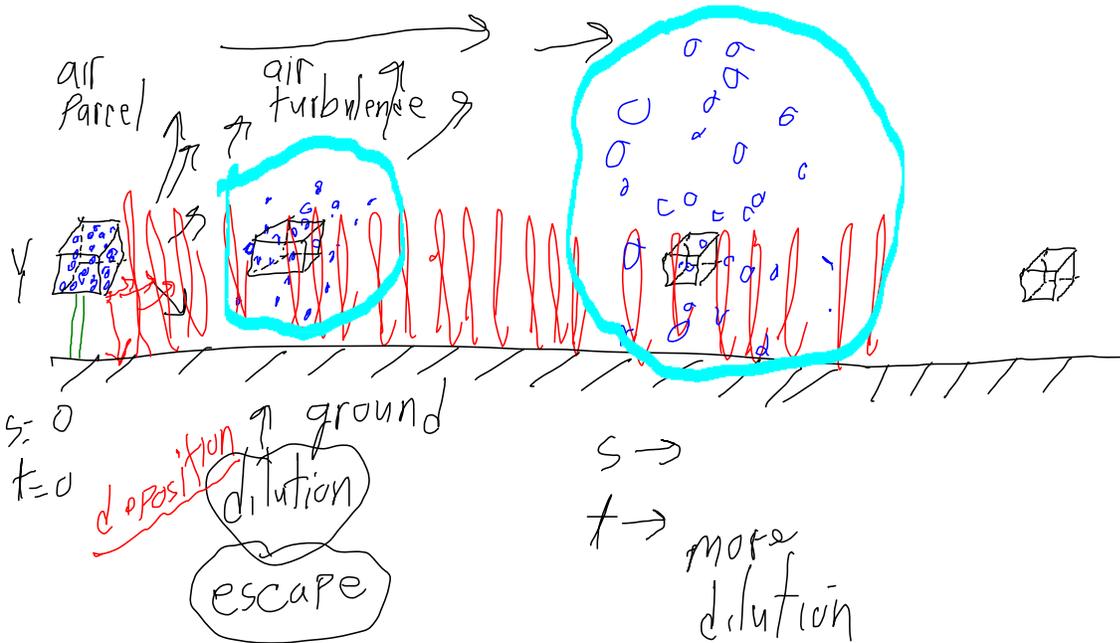
Spore **dilution & escape** from canopy
(high air turbulence)

- *high plant density* *low plant density*
- Other models have been proposed, including combination of exponential and power functions (for mixture of deposition and dilution/escape). These two are sufficient for many studies

$$Y = A e^{-b_E s - b_P s}$$

See: Aylor (1999) -- citation in book;
Also: Reynolds (Dec. 2011). *Phytopathology* 101: 1465-70.

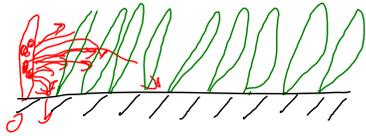
splash dispersal





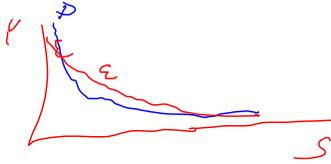
high turbulence

dilated escape



low air turbulence

deposition



Use of models

- **Model selection**
 - **Graphical and statistical approaches** (as with disease progress curves)
 - **Plots of dY/ds (estimated) versus s , Y versus s ; and $\ln(Y):s$ and $\ln(Y):\ln(s+\lambda)$ $\lambda=0$ ← best way**
 - Unlike the case with temporal analysis, Y^* is the same for the two models, but s^* is not
 - Either s or $\ln(s+\lambda)$
- **Model fitting**
 - Ordinary linear least squares
 - $\ln(Y)$ versus s or $\ln(Y)$ versus $\ln(s+\lambda)$
 - ◆ Use $\lambda=0$ (standard)
 - ◆ Or, try several λ values, from 0 up to the shortest observed distance
 - ◆ Analogous to trying different models
 - ◆ My preference: 1/2 between 0 and first distance
 - Nonlinear least squares (or maximum likelihood or Bayesian)
 - Can directly estimate λ
 - Evaluation of **residuals**

$$\frac{dY}{ds} = \frac{\Delta Y}{\Delta s}$$

s	Y
10	50
20	25
30	10
40	5

50-25
20-10
25-10
30-20

$$Y^* = a \cdot s - b \cdot \lambda^* \cdot s$$

$$\ln(Y) = a \cdot s - b \cdot \lambda^* \cdot s$$

$$\ln(Y) = a \cdot s - b \cdot \lambda^* \cdot \ln(s + \lambda)$$

Predictor

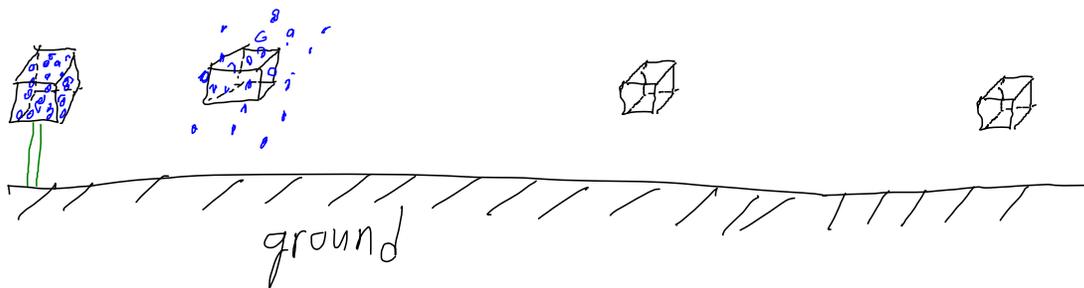
$$\ln(Y) \begin{cases} \ln(s+0) \\ \ln(s+1) \\ \ln(s+2) \\ \vdots \\ \ln(s+7.99) \end{cases}$$

Reading assignment:

Chapter 7

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