

# A Multi-Channel Passive Brain Implant for Wireless Neuropotential Monitoring

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**Abstract**—We propose a novel multi-channel (8 channels) passive neuro-sensing system for wireless acquisition of brain signals as low as  $20 \mu V_{pp}$ . Compared to previous batteryless multi-channel neuropotential sensors, the proposed design exhibits: a) 28 times better sensitivity, b)  $\sim 2$  times smaller footprint, and c) scalability to 100s or even 1000s of channels. The proposed system consists of an external interrogator and a neuro-recorder implanted inside the scalp. For operation, the interrogator sends a) a 2.4 GHz carrier signal to “turn on” the implant, and b) an infrared control signal for channel selection. The latter activates the desired channel via a photo-activated multiplexer. For this channel, the carrier signal is mixed with the neural signal ( $f_{neuro}$ ) to generate a  $4.8 GHz \pm f_{neuro}$  modulated signal. The latter is then transmitted back to the interrogator. To verify the implant’s operation inside biological tissues, *in-vitro* measurements are presented using pig skin. Experimental results show that the proposed neuropotential recorder exhibits  $20 \mu V_{pp}$  sensitivity at all eight channels (viz. it can record any signal generated by the human brain). The system is also in compliance with the strictest Federal Communications Commission standards for patient safety. Notably, the proposed approach is scalable to a much higher number of channels. As such, the proposed system can be a game-changing capability for a wide range of applications.

**Keywords**—Brain implant, wireless, multichannel, neurosensing, passive, infrared photodiode, photovoltaic cell, antiparallel diode pair, biocompatibility, biomedical telemetry.

## I. INTRODUCTION

DEEP brain neuropotential monitoring can significantly improve the individual’s physical and mental well-being. Example applications include: 1) detection and interruption of early epileptic seizures, 2) behavioral studies to determine levels of consciousness, 3) understanding and improving the brain’s functionality for patients with Alzheimer’s and Parkinson’s conditions, among others [1], [2].

Current technology on deep brain neuropotential recording is limited by several factors. Specifically, wires are typically used to connect the implant recorder to external monitoring units [2], [3]. These tethered connections are obtrusive to the patient and restrict long-term brain clinical research in free-moving subjects. To address this concern, wireless brain implants have been pursued. However, current versions of these implants are typically bulky and use batteries or harvesters for powering them [4]. Also, such implants are associated with temperature increases, implying possible damage to the brain tissues or cells [5]. To address these concerns, a batteryless wireless neuropotential recorder was proposed in [6]. However, the recorder still required a battery that was placed external to the skull, limiting comfort, and increasing the risk of infection. Notably, wirelessly powered neural recorders have been reported in [7]–[11], but these still employ complex integrated circuits.

As an alternative, we recently introduced a new class of wireless fully-passive brain implants [13]–[16] that use simple

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TABLE I  
VOLTAGE AND FREQUENCY RANGE OF SIGNALS GENERATED BY THE HUMAN BRAIN [12]

Neural Signals	Voltage Range	Frequency Range
Local field potentials	$20 \sim 2000 \mu V_{pp}$	10~200 Hz
Neural spikes	$100 \mu V_{pp}$	300 Hz~5 kHz

electronic circuitry. In our latest work [16], we demonstrated a wireless and fully-passive brain implant, but it was used only for single-channel recording. This implanted recorder exhibited a very small footprint of  $8.7 \text{ mm} \times 10 \text{ mm}$  and was shown to exhibit sensitivity as high as  $20 \mu V_{pp}$ . As such, all signals generated by the human brain, see Table I [12], can be detected. However, the single channel operation of our earlier device [16] is not suited for realistic clinical applications. For the latter, we require 100 or even 1000s of channels of concurrent neuropotential recording.

In this paper, we propose multi-channel neuropotential recording that is wireless and batteryless. Specifically, we design and fabricate a proof-of-concept wireless multi-channel neuropotential monitoring system with the following unique features:

- 1) wireless, passive and biocompatible operation,
- 2) concurrent recording from 8 channels by using an infrared transceiver/receiver, scalable to 100s and up to 1000s of channels,
- 3) neuropotential detection as low as  $20 \mu V_{pp}$  per channel,
- 4) validation using fresh pig skin to emulate the human skin’s properties at RF and infrared bands,
- 5) extremely small power consumption ( $98 \mu W$ ),
- 6) compliance with the strictest Federal Communications Commission (FCC) standards for patient safety [17].

As compared to previous batteryless multi-channel neuropotential recordings [18], the proposed implant exhibits: a) 28 times higher sensitivity, b)  $\sim 2$  times smaller footprint (1600  $mm^2$  vs 3000  $mm^2$  in [18]), c) higher scalability (3 photodiodes for 8 channels vs. 8 photodiodes for 8 channels in [18]), and d) alternative and, likely, more practical implementation (using infrared instead of visible light as in [18] for improved skin penetration).

Unfortunately, the design in [16] refers to a single channel and is not scalable for the realistic requirements of 100s and 1000s channel recordings. These multi-channel recordings are needed to understand movement of multiple muscles and study how chronic issues such as Parkinson's and Alzheimer's affect brain functionality.

A number of innovations are brought forward to achieve the proposed scalable multi-channel capabilities, including:

- 1) use of an infrared exterior transceiver and implanted receiver,
- 2) integration of an implanted multiplexer,
- 3) re-design/optimization of the implanted and interrogator antennas for optimal coupling to the new implanted multiplexing unit,
- 4) re-design/optimization of the entire brain implant and exterior interrogator to accommodate these new features, and
- 5) use of actual pig-skin, rather than a tissue-emulating phantom, to achieve the desired performance at optical frequencies.

## II. NEUROSENSING SYSTEM OVERVIEW

The block diagram of the proposed neuropotential transceiver and receiver is depicted in Fig. 1. The set-up consists of 1) an implanted recorder placed just under the scalp with the recording electrodes protruding through the bone into the brain, and 2) an external interrogator placed outside the scalp. This operation of the proposed neurosensor involves two processes: 1) wireless monitoring of the neural signal using a process similar to RFIDs, 2) selection/toggle of the different channels via an infrared-enabled implanted multiplexer.

The process for wireless and batteryless neural signal collection/monitoring can be summarized as follows. First, the interrogator sends a 2.4 GHz carrier signal to "turn on" the implanted recorder. The mixer in the implanted device uses the 2.4 GHz signal to generate a  $4.8 \text{ GHz} \pm f_{neuro}$  modulated signal that is eventually transmitted back to the interrogator. To obtain low conversion loss and suppress the DC term in the mixing process, an anti-parallel diode pair (ADPD) configuration is employed, as described in [14].

For toggling different channels, an extremely low-power commercial multiplexer (Analog Devices, ADG708) is adopted. Further, to provide power and control to the multiplexer, we employed an implanted infrared receiver and an exterior infrared transceiver as shown in Fig. 2. For this particular multiplexer, each of the 8 channels is selected via an "on-off" scheme that employs 3 implanted photodiodes (Vishay, VEMD2000). These three photodiodes can be individually turned on and off to generate 8 different optical combinations. The optical signal is denoted as  $P_0$ ,  $P_1$  and  $P_2$  as depicted in Fig. 2. Each of the 8 combinations is then

used via the multiplexer to select one of the 8 probes for recording. Notably, generation of this  $P_0P_1P_2$  code is done via the external infrared emitters as in Fig. 1 and 2. These emitters also pass their signal through the skin to the photovoltaic cell to introduce a stable DC voltage on the multiplexer of the implant. It is noted that thereby power consumption of the implant is due to the multiplexer operation and the resistance of the photodiodes. But the power consumption of the multiplexer operation can be reduced to trivial level by choosing high-value resistors and high-sensitivity photodiodes. Typically, when the series resistance of the photodiode is  $R = 1.5M \Omega$ , see Fig. 2, the power consumption is only  $16 \mu W$  ( $-18 \text{ dBm}$ ).

In designing the aforementioned neuropotential monitoring system, the goal is to achieve  $20 \mu V_{pp}$  sensitivity for all channels. This would enable recording of all possible neural signals generated by the human brain as shown in Table I. Typically, the minimum detectable neuropotential signal level can be expressed as [16]:

$$MDS_{neuro}[\text{dBm}] = \text{Receiver Sensitivity}[\text{dBm}] + L_{sys}[\text{dBm}] \quad (1)$$

where  $L_{sys}$  = overall system loss, and in the above  $\text{Receiver Sensitivity}$  = minimum detectable signal level by the receiver. For our employed interrogator, the sensitivity is  $-120 \text{ dBm}$  [15]. Thus, to guarantee a minimum detectable signal of  $20 \mu V_{pp}$  (or  $-90 \text{ dBm}$ ),  $L_{sys} < 30 \text{ dB}$ . The receiver system loss,  $L_{sys}$ , could be divided into three major components as expressed in (2):

$$L_{sys}[\text{dB}] = L_{prop}[\text{dB}] + L_{conv}[\text{dB}] + L_{match}[\text{dB}] \quad (2)$$

In the above,  $L_{prop}$  = propagation loss between the implanted and the interrogator antenna at  $4.8 \text{ GHz} \pm f_{neuro}$ ,  $L_{conv}$  = conversion loss at the implanted mixer, and  $L_{match}$  = impedance mismatch losses between the antenna and the mixer in the implanted device. To ensure  $L_{sys} < 30 \text{ dB}$ ,  $L_{prop}$ ,  $L_{conv}$  and  $L_{match}$  should be minimized to the extent possible. This is discussed in Section III.

Table II compares the proposed multichannel neuropotential monitoring system with previously reported wireless and batteryless neural recorders. The neural recorders in [13]–[16] have only one channel and are therefore not sufficient for realistic clinical applications. A batteryless multichannel system was reported in [18] but this implant has  $\sim 2$  times larger footprint to achieve 28 times lower sensitivity. Also, in [18], visible light was employed to excite the photodiodes. But visible light cannot penetrate the biological tissues. Further, the design in [18] exhibited limited scalability, requiring 8 photodiodes for 8 channels (rather than 3 photodiodes as done here).

In this paper, we propose the first wireless, passive and scalable fully-implanted device with  $20 \mu V_{pp}$  sensitivity at all channels. Infrared light is used to penetrate through tissues and toggle an N-input multiplexer to activate  $2^N$  channels.

For this paper, the focus was totally on the multichannel implant design and demonstrations. Similarly, the implant's

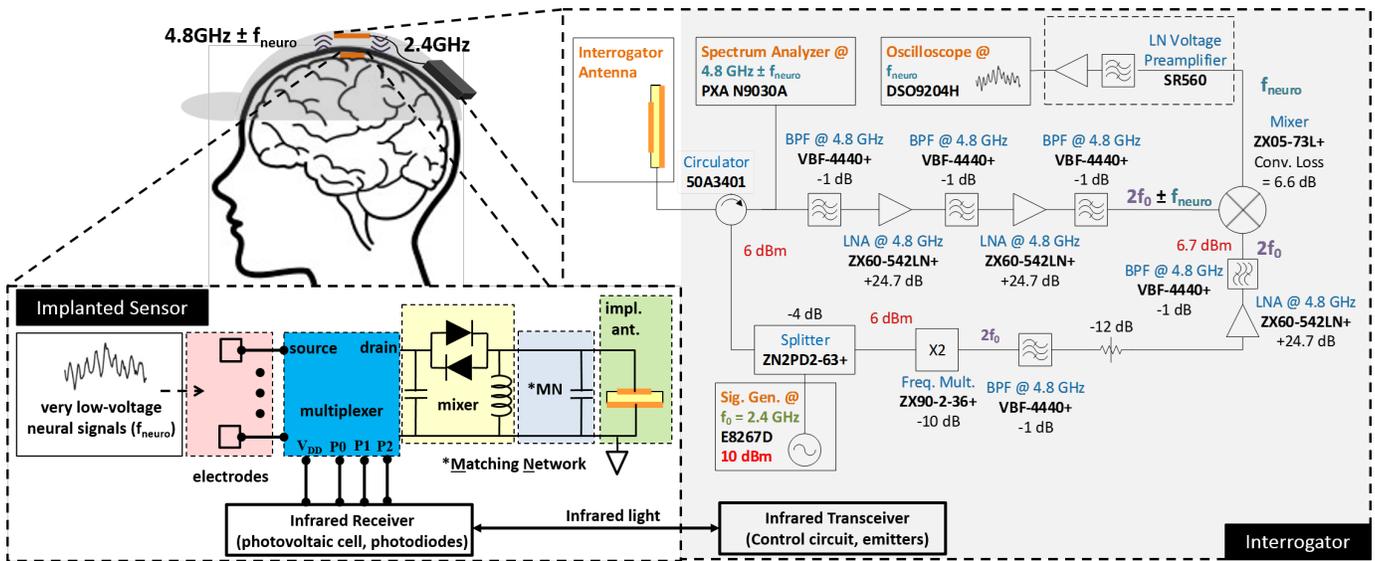


Fig. 1. Block diagram of the proposed multichannel neurosensing system.

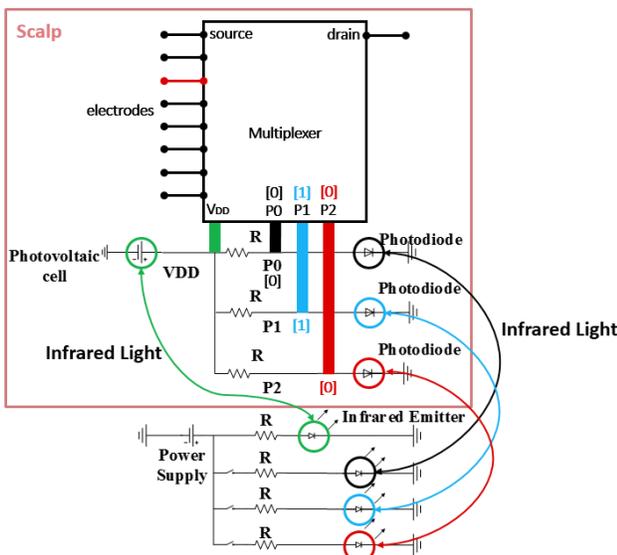


Fig. 2. Proposed implanted infrared receiver and exterior transceiver used to toggle different neuro-channels in a wireless and passive manner. The transceiver employs 3 photodiodes to form a 3 digit code for selecting each of the 8 channels. In this condition, the three digit code is 0,1,0 and channel 3 is selected.

power consumption can be further reduced in the future by using higher-permittivity and lower-loss substrates. The power consumption in Table II shows the average power of all 8 different channel combinations (from 0,0,0 to 1,1,1).

### III. BRAIN IMPLANT AND INTERROGATOR DESIGN

The developed brain implant and interrogator PCB layouts are shown in Fig. 3. For a proof-of-concept demonstration, the implant and interrogator each employed a three-layer metallization structure, occupying a footprint of  $40\text{ mm} \times 40\text{ mm}$  in the area. Specifically, for the implanted neural recorder,

the photovoltaic cells and the implanted antenna were placed on the top layer to receive the RF and infrared signals from the interrogator. The multiplexer and mixing circuits were located on the bottom layer. The middle layer contains the common ground shared by the antenna, multiplexer, and the mixing circuits. A similar structure was employed for the interrogator's circuit, as shown in Fig. 3(b). Co-simulations were performed in ANSYS HFSS and Keysight ADS. An in-house genetic algorithm was also employed for design optimization [19]. Below, we discuss the various components that comprise the transceiver.

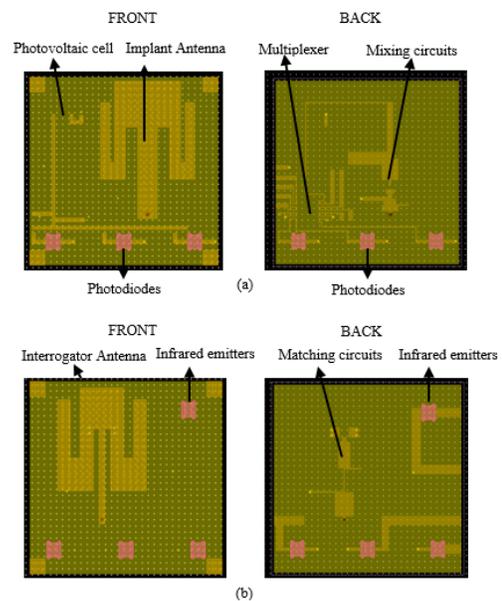


Fig. 3. Proposed system layout: (a) brain implant, and (b) exterior interrogator. The corresponding areas both are  $40\text{ mm} \times 40\text{ mm}$

TABLE II  
COMPARISON BETWEEN PROPOSED VS PREVIOUSLY WIRELESS IMPLANTED DEVICES WITHOUT INTEGRATED CIRCUITS

Ref	Type	Footprint	Power consumption	Channels	Operation distance	Operation frequency	Min. detectable signal
[13]	Implanted	$12 \times 4 \text{ mm}^2$	N/A	1	$< 1.5 \text{ cm}$	2.4 GHz and 4.8 GHz	$3.4m V_{pp}$ (in-vitro)
[14]	Implanted	$39 \times 15 \text{ mm}^2$	0.008 mW	1	8 mm	2.4 GHz and 4.8 GHz	$200 \mu V_{pp}$ (in-vitro)
[15]	Implanted	$16 \times 15 \text{ mm}^2$	0.035 mW	1	$\sim 1.5 \text{ cm}$	2.4 GHz and 4.8 GHz	$63 \mu V_{pp}$ (in-vitro)
[16]	Implanted	$8.7 \times 10 \text{ mm}^2$	0.305 mW	1	2 mm	2.4 GHz and 4.8 GHz	$20 \mu V_{pp}$ (in-vitro)
[18]	Implanted	$\sim 50 \times 60 \text{ mm}^2$	N/A	3	3 mm	2.45 GHz and 4.9GHz	$700 \mu V_{pp}$ (in-vitro)
Proposed	Implanted	$40 \times 40 \text{ mm}^2$	0.598 mW	8	2.5 mm	2.4 GHz and 4.8 GHz	$20 \mu V_{pp}$ (in-vitro)

### A. Implanted and Interrogator Antennas

The implants in [13] and [15] were designed to operate under the skull, while the implants in [16] and this work are intended for operation under the skin. Our target distance of 2.5 mm was selected according to the thickness of the pig-skin used in this experiments. This aligns well with real-life applications, where the thickness of the human head skin ranges from 2 mm to 4 mm, according to age [20]. In this paper, the implanted and interrogator antennas have been optimized for the aforementioned 2.5mm distance.

The implanted and interrogator antennas were designed to overcome propagation loss at 2.4 GHz and  $4.8 \text{ GHz} \pm f_{neuro}$  and ensure FCC regulations [17]. To achieve dual band radiation, an E-shaped patch antenna geometry was adopted for the implanted and interrogator antennas, shown in Fig. 3. The simulated transmission coefficient  $|S_{21}|$  between the implanted and interrogator antennas as a function of the depth into the skin tissue is given in Fig. 4. As seen,  $|S_{21}| = -6\text{dB} - 14\text{dB}$  at 2.4 GHz/4.8 GHz when the implant is 1 mm below the skin. Notably, our interrogator/implanted antenna pair is designed and optimized in the reactive near-field mode. Because of the poor antenna gain in the far-field mode, any unintended 2.4 GHz signal is precluded from reaching the implant. Even if a strong interference 2.4 GHz signal was present, the power would not be sufficient to activate the brain implant given the poor antenna gain and tissue loss.

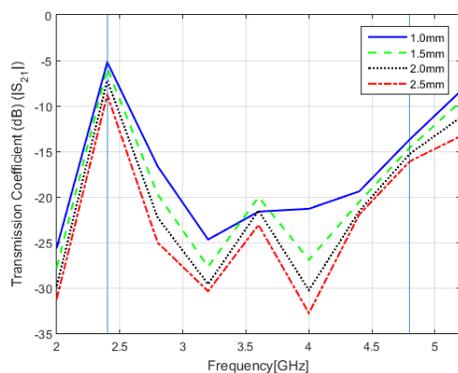


Fig. 4. Simulated transmission coefficient,  $|S_{21}|$ , between the transmitting and receiving antennas for different pig skin depth.

### B. Implant Mixer

The goal of the mixer design is to decrease the conversion and mismatch losses ( $L_{conv}$  and  $L_{match}$ ) mentioned in Section

II. The employed mixer consists of the following:

- 1) Antiparallel Diode Pair (APDP),
- 2) Inductor that provides the circuit's route to ground for the low-frequency neuropotential signal ( $f_{neuro}$ ),
- 3) Capacitor that acts as short for the 2.4GHz to isolate the DC from the neuropotential signal ( $f_{neuro}$ ),
- 4) Matching circuit to minimize mismatch losses between the antenna ( $50 \Omega$ ) and mixer. To further reduce the number of lumped elements, the inductor and capacitor were replaced by short- and open-circuited transmission lines, respectively. The final mixer and matching circuit PCB layout is shown in Fig. 3(a). It is noted that commercial APDP diodes (Avago Technologies, HSMS-286C) with good circuit balance were employed for subharmonic mixing.

### C. Infrared Transceiver/Receiver

The infrared exterior transceiver and implanted receiver are intended to provide 1) stable DC voltage of 3V, and 2) a control signal to the implanted multiplexer for toggling among the different channels. To ensure batteryless operation, the DC voltage is provided via an infrared-illuminated photovoltaic cell. The control signal is provided by the 3 digit code generated by the photodiodes. It is important that the infrared receiver have sufficient isolation to ensure detection of the 3 digit code. Otherwise, the infrared emitter can lead to erroneous channel turn on. To avoid this, highly directive photodiodes and photoemitters were selected (The patterns of the photodiodes are given in Fig. 5(a) and 5(b)).

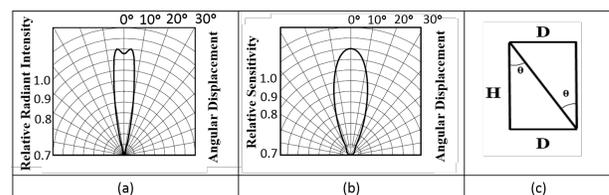


Fig. 5. Radiation and reception pattern for the (a) infrared emitter, (b) photodiodes and (c) the geometry of the photodiodes and photo emitters

As seen in Fig. 5, the radiation receiving sensitivity at  $20^\circ$  from the normal intensity is rather low. This ensures excellent isolation between the adjacent photodiodes. The suggested separation between two photodiodes should be:

$$D = H \cdot \tan\theta \quad (3)$$

where  $H$  = thickness of the skin, and  $\theta$  = infrared beam angle between the photodiode and the neighboring emitter. Typically,  $5 \text{ mm} < H < 8 \text{ mm}$  and  $\theta < 20^\circ$ . Based on these averages, the distance between the photodiodes should be at least  $D \approx 2 \text{ mm}$ . That is, the goal isolation between different channels can be achieved by the designated physical placement of the photodiodes.

#### D. RF Interrogator

The RF interrogator is intended to: a) generate and receive the backscattered  $4.8 \text{ GHz} \pm f_{neuro}$  after third-order mixing, and b) demodulate the mixing product and display/store the recovered time domain signal. As would be expected, demodulation can be performed by mixing the backscattered signal with another 4.8 GHz reference signal to obtain the "baseband" neuropotential at  $f_{neuro}$ .

The employed interrogator system is displayed in Fig. 1 and a similar version was used in [16]. For the set up in Fig. 1, multiple stages of filtering and amplification were used to improve the signal to noise ratio. Notably, this architecture has a very low noise figure of 3.8 dB.

### IV. NEUROSENSING SYSTEM PERFORMANCE

#### A. Fabricated Prototype

The fabricated brain implant and interrogator prototypes are shown in Fig. 6. As mentioned earlier, for this proof-of-concept demonstration, both devices occupy a footprint of  $40 \text{ mm} \times 40 \text{ mm}$ . They were fabricated on Rogers RO4003C substrate ( $\epsilon_r = 3.38$ ,  $\tan\delta = 0.0021$ ) of thickness 40 mils (1.016 mm). To ensure biocompatibility, the implanted recorder was coated with a 0.7 mm-thick layer of Polydimethylsiloxane ( $\epsilon_r = 2.8$ ,  $\tan\delta = 0.001$  [15]). This relatively lossless layer decreases the power absorbed by the human tissue, thereby reducing losses between the implant and interrogator antennas [21].

Selection of a higher permittivity substrate will further miniaturize the implant by shrinking the antenna and microstrip feed lines. We have already identified high permittivity materials (TMM 13i,  $\epsilon_r = 12.2$ ,  $\tan\delta = 0.0019$ ) that exhibit the similar loss tangent to the low permittivity dielectric employed in this work (RO4003C,  $\epsilon_r = 3.38$ ,  $\tan\delta = 0.0021$ ). We have also identified flexible polymer-ceramic composites having  $\tan\delta \approx 0.0025$  for  $\epsilon_r \approx 10$  [22]. That is, we are confident that we can switch to higher-permittivity dielectrics without increasing dielectric losses.

#### B. Measurement Setup

The measurement setup used to validate the  $20 \mu\text{V}_{pp}$  sensitivity of our neuropotential detector for all channels is shown in Fig. 7. As depicted, the 2.4 GHz carrier was a pure 6 dBm sinusoid that was supplied to the interrogator using a signal generator (Agilent SG386). Also, the emulated neuropotentials at  $f_{neuro} = 10 \text{ Hz}$  to  $5 \text{ kHz}$  were generated using an arbitrary function generator (Leader, LFG-1300). For the infrared control circuit, shown in Fig. 6, manual turn on/off switches were used. To emulate the human head

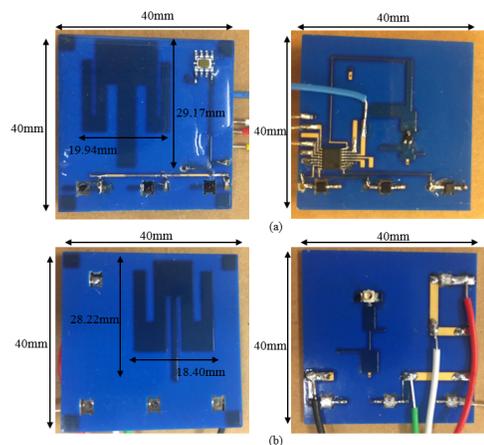


Fig. 6. Fabricated prototypes: (a) brain implant, (b) exterior interrogator

tissues, the neural recorder was immersed inside a four-layer head phantom shown in Fig. 6. For this phantom, the bone, dura/gray matter, and white matter tissues were formulated using the recipes in [23]. For the skin, we used fresh pig skin layer [24]. Conventional phantoms accurately emulate the dielectric properties of human tissues, but not their light penetration properties. With this in mind, pig skin (viz. actual biological tissue) is selected as to better emulate both RF and infrared signal propagation. The permittivity and loss tangent of the pig-skin were measured using the Agilent 85070E Dielectric Probe Kit and further compared vs. the theoretical skin properties [25] shown in Fig. 8. The reader is referred to [26]–[28] for infrared losses through the scalp.

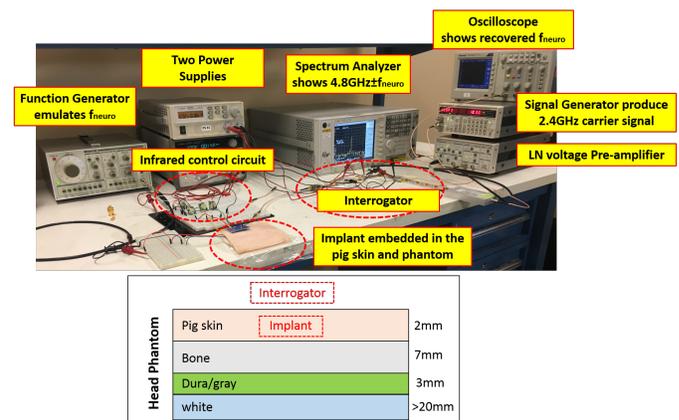


Fig. 7. The neurosensing measurement setup with the layered head phantom.

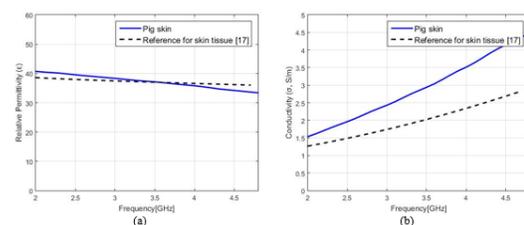


Fig. 8. (a) Measured permittivity, and (b) measured conductivity of pig-skin versus the reference skin properties reported in [25].

### C. Minimum detectable signal

As already noted, our goal is to detect neural signals down to  $20 \mu V_{pp}$ . To achieve this, we must minimize the overall losses between the interrogator and implant. This overall system loss,  $L_{sys}$ , is defined as the difference between the power of the neuropotential signals detected at the input of the implanted device ( $f_{neuro}$ ) and the backscattered power received at the interrogator ( $4.8 \text{ GHz} \pm f_{neuro}$ ). Based on simulations, we found that  $L_{sys} = -18.2 \text{ dB}$  when  $10 \text{ Hz} < f_{neuro} < 5 \text{ kHz}$  for a distance of 2.5 mm between the implant and interrogator. Our measurements using a pig skin thickness of 2.0 mm indicated that  $L_{sys} = -25 \text{ dB}$ . Although  $L_{sys}$  is 7 dB larger than simulation, it still meets the system loss criteria of  $L_{sys} < 30 \text{ dB}$ . Fig. 9 shows example demodulated backscattered signals received at the interrogator. For these example waveforms, the emulated neuropotentials were as low as  $20 \mu V_{pp}$ . The observed higher noise can be attributed to the preamplifier filter used at the interrogator. This noise can be suppressed by using filters.

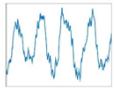
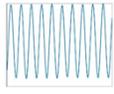
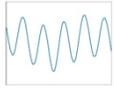
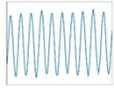
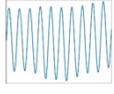
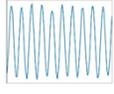
$f_{neuro}$ (Hz)	$f_i$ and $f_{fit}$ (Hz)	Recovered Waveform	$f_{neuro}$ (Hz)	$f_i$ and $f_{fit}$ (Hz)	Recovered Waveform
10	1-10		500	300-1000	
50	10-300		1000	300-1000	
100	10-300		5000	3K-10K	

Fig. 9. Time-domain performance for minimum detectable neural signal ( $MDS_{neuro} = 20 \mu V_{pp}$ )

### D. Specific Absorption Rate (SAR)

To confirm that the proposed neurosensing system meets the FCC safety guidelines, a 10-cm-radius spherical head model was employed. This model was composed of skin (9.2-cm-radius), bone (8.5-cm-radius), gray matter (7.8-cm-radius), and white matter (7.5-cm-radius) tissues [29]. The implant was placed in the middle of the skin layer.

For SAR simulation, the brain implant was placed below the skin layer and the interrogator was placed right above it. It was found that the average SAR for 1 g of tissue with 6 dBm carrier power at 2.4 GHz had a max value of  $SAR_{1g} = 0.368 \text{ W/kg}$ . This value conforms to the strictest FCC requirements of  $SAR_{1g} < 1.6 \text{ W/kg}$  for uncontrolled environment exposure [17].

## V. CONCLUSION

A multichannel passive neurosensing system was proposed for the wireless acquisition of deep brain signals as low as  $20 \mu V_{pp}$ . This multichannel neurosensors employed an implanted multiplexer and an infrared transceiver/receiver module. An 8-channel prototype was designed and demonstrated to achieve  $20 \mu V_{pp}$  sensitivity in all channels under *in-vitro* conditions.

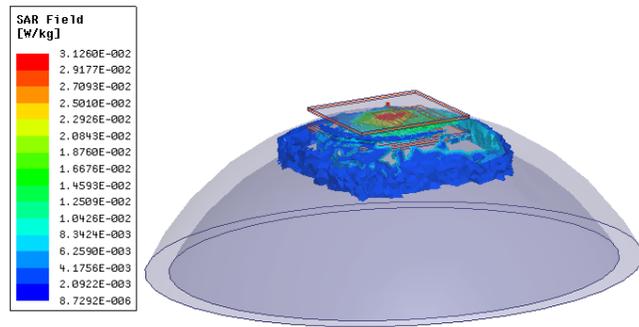


Fig. 10. SAR performance averaged over 1 g with input carrier power at 2.4 GHz and 6 dBm.

That is, most neural signals generated by the human brain can be detected across all 8 channels in a passive and wireless manner. This is a game-changing capability for a very wide range of applications.

Future work will target three major directions:

1) In-vivo testing. The ultimate goal of the wireless passive implanted system is to record neuropotentials *in-vivo*. In the past, we successfully recovered ECG (Electrocardiography) signals from the human body using commercial electrodes attached to our device. Future studies will explore brain signal acquisition from animals. We note that *in-vivo* data are expected to be noisier (e.g., motion artifacts or noise from other equipment) and can be possibly corrupted by 50/60 Hz signals associated from power lines in the operation room. In this case, notch filtering will be required to remove the 50/60 Hz interference, and more sophisticated signal processing techniques may be needed to retrieve the neural signals (e.g., window averaging).

2) System miniaturization. For our proof-of-concept multi-channel board reported in this work, the footprint of the circuit is relatively large ( $40 \text{ mm} \times 40 \text{ mm}$ ). The largest portion of the circuit area is occupied by the implanted antenna. Therefore antenna miniaturization will be explored by: 1) choosing higher dielectric constant substrates, and 2) employing more sophisticated antenna designs.

3) Channel scalability. In our proposed system, channel scalability is achieved by using N photodiodes for  $2^N$  channels. For the 8-channel system reported in this work, 3 photodiodes are employed. Using 10 photodiodes, we can reach  $2^{10} = 1024$  channels. To maintain a small implant size, customized photodiodes and multiplexers would also be required. The former implies frequency-dependent photodiodes that use both frequency and physical separation to reduce total footprint. We would also require miniature in-house multiplexer designs. Higher permittivity materials can be employed to reduce the antenna footprint and microstrip line size. In terms of power consumption, if the photodiode serial resistance is kept high ( $M\Omega$  level, per Section II of the manuscript), it is predicted that the estimated power consumption by a 10-photodiode layout will be similar to that of the 3-photodiode layout reported in this work.

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