Enhancing Plant Resistance at the Seed Stage: Low Concentrations of Methyl Jasmonate Reduce the Performance of the Leaf Miner *Tuta absoluta* but do not Alter the Behavior of its Predator *Chrysoperla externa*

Priscila Strapasson • Delia M. Pinto-Zevallos • Sulav Paudel • Edwin G. Rajotte • Gary W. Felton • Paulo H. G. Zarbin

Received: 11 April 2014/Revised: 25 June 2014/Accepted: 27 August 2014/Published online: 16 October 2014 © Springer Science+Business Media New York 2014

Abstract Plants express inducible direct and indirect defenses in response to herbivory. The plant hormone jasmonic acid (JA) and related signaling compounds referred to as jasmonates play a central role in regulating defense responses to a wide range of herbivores. We assessed whether treating tomato seeds with 0.8 mM of methyl jasmonate (MeJA) affected the performance of the leaf miner Tuta absoluta, and whether possible changes in volatile profiles altered the behavior of its predator Chrysoperla externa. MeJA-treatment significantly lengthened larval development and decreased the pupal weight of T. absoluta. Herbivory alone increased the emissions of α -pinene, 6-methyl 5-hepten-2-one, β -myrcene, (E)- β -ocimene, isoterpinolene, TMTT, (Z)- β -hexenyl butyrate, and hexyl salicylate. MeJA seed treatment significantly decreased the emissions of α -cubebene from undamaged and herbivore-infested plants. In addition, the emissions of several compounds were lower in the absence of herbivory. Chrysoperla. externa preferred odors from herbivoreinfested plants over those from control plants, regardless of the MeJA-treatment, and they did not show any preference for herbivore-infested plants for any of the MeJA-treatments. Our results show preliminary evidence that the treatment of tomato seeds with MeJA can reduce the performance of Tuta absoluta, and that the chemical differences observed in plant VOC profiles do not alter the behavior of the model predator.

P. Strapasson • D. M. Pinto-Zevallos • P. H. G. Zarbin (⊠) Laboratory of Semiochemicals, Department of Chemistry, Federal University of Paraná, P.O. Box 19081, C.E.P, 81531-980 Curitiba, Paraná, Brazil e-mail: pzarbin@quimica.ufpr.br

S. Paudel · E. G. Rajotte · G. W. Felton Department of Entomology, Pennsylvania State University, University Park, PA 16802, USA Keywords Methyl jasmonate · Pest resistance · Volatile organic compounds · Plant-insect interactions · Pest management · Lepidoptera · Gelechiidae · Neuroptera · Chrysopidae

Introduction

Plants have evolved an array of mechanisms to defend themselves against the hostile biotic environment. In addition to constitutive defenses, they can express inducible defenses that are activated in response to adverse conditions such as herbivory (Karban 2011; Karban and Baldwin 1997) that include the increase of secondary metabolites (Mithöfer and Boland 2012) and the induction of structures such as trichomes (Traw and Dawson 2002), which will directly impact the attacking herbivore. Furthermore, inducible defenses include qualitative and/or quantitative changes in the constitutive emissions of volatile organic compounds (VOCs), which are known to attract natural enemies (predators and parasitoids) that provide biological control of herbivores (Pinto-Zevallos et al. 2013; Turlings and Wäckers 2004). The expression of herbivoreinduced defenses is mediated primarily by the phytohormones jasmonic acid (JA), ethylene, and salicylic acid (SA) (Smith et al. 2009; Wu and Baldwin 2009). The role of JA has been demonstrated in the induction of several defense traits including secondary metabolites (Van Dam et al. 2004), defensive proteins such as polyphenol oxidase and proteinase inhibitors (Farmer and Ryan 1992; Felton et al. 1989), trichomes (Boughton et al. 2005), and plant VOCs (Ament et al. 2004; van Poecke and Dicke 2002). JA-mediated defenses are particularly activated in response to chewing insects such as lepidopteran caterpillars (Kessler and Baldwin 2002).

The potential for enhancing plant resistance by using selected chemical molecules (plant elicitors) has been known for several years (Conrath et al. 2006) and may be a good strategy to be incorporated in pest management programs (Pinto-Zevallos and Zarbin 2013; Stout et al. 2002). Induction of resistance at the seed stage by using chemical elicitors, including phytohormones and their derivates, improves the resistance of the plants to adverse conditions of temperature (Farooq et al. 2008), salinity (Shakirova et al. 2003), water stress (Li et al. 1998), heavy metals (Krantev et al. 2008), and to biotic stresses, particularly microorganisms (Buzi et al. 2004; Pankaj et al. 2013). However, evidence of the potential of seed treatments for improving resistance to arthropod pests is scarce. It recently has been found that treating tomato seeds with JA not only has detrimental effects on herbivores of multiple arthropod taxa, but is also beneficial to the plant since the leaf area grazed by caterpillars was reduced (Worrall et al. 2012).

The tomato leafminer or pinworm Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) is an oligophagous herbivore that feeds on Solanaceae species. In its native range of South America, it is considered the major pest of tomato (Villas Bôas et al. 2009). Currently, it is threatening tomato production in Europe where it was introduced a few years ago, and is rapidly spreading towards the Middle East, and North and West Africa (Tuta absoluta Information Network 2014). In the absence of effective alternative strategies, multiple applications of chemical pesticides (10-30 application/crop cycle) are used to manage T. absoluta (Sigueira et al. 2000). Continuous overuse of pesticides can result in environmental contamination, adverse effects on non-target organisms (El-Wakeil et al. 2013), and rapid development of resistance (Lietti et al. 2005; Silva et al. 2011; Siqueira et al. 2000). Thus, the use of the plants' own defense mechanisms to induce resistance against insect pests may play a role, as a component of an integrated pest management (IPM) approach, in reducing pest damage while also minimizing the adverse effects of pesticide use.

The aim of this study was to assess whether treating the tomato seeds with methyl jasmonate (MeJA) can confer long lasting resistance of tomato plants against T. absoluta. Additionally, we assessed whether the MeJA treatment can alter the induction of volatile organic compounds upon herbivory, and whether the foraging behavior of a natural enemy of T. absoluta is affected. We chose Chrysoperla externa (Neuroptera: Chrysopidae) as the model predator. Immature stages of this species are known as voracious consumers of various herbivores including eggs and young larvae of T. absoluta (Ghoneim 2014). Chrysoperla externa has been regarded as a good candidate to be incorporated in biological control programs. Mass rearing of this species is easy, and it has already proven its potential as a biological control agent of T. absoluta (Embrapa 2014; Ghoneim 2014).

Methods and Materials

Living Material Larvae and pupae of Tuta absoluta were collected from a commercial organic tomato cultivar Cereja (cherry tomato) farm in Curitiba (Paraná, South Brazil; (25° 17' 31" S, 49° 13' 26" W) and were further maintained on tomato cultivar Santa Clara in controlled conditions (20± 1 °C; 70 ± 10 % RH; L:D 12:12 h) at the Department of Biological Sciences from the Federal University of Paraná. The rearing of Chrysoperla externa was started from eggs of a colony established at the Federal University of Lavras (Lavras, Minas Gerais, Brazil), and maintained at 25 °C and a photoperiod of 14:10 h (L:D). Larvae were fed on frozen eggs of Anagasta kuehniella until pupation. Adults were kept in a PVC cage with the interior wall lined with paper to support oviposition. Adults were provided with an aqueous solution of honey and yeast. After seed treatment, commercial tomato seeds cv. Santa Clara were sown in pots filled with a commercial substrate (Tropstrato HA, Vida Verde, São Paulo, Brazil). Two to three weeks after plant emergence, they were transferred to individual pots (volume ca. 150 ml). Seedlings were kept at room temperature with artificial lights (L:D cvcle 14:10 h). They were watered every 3-4 days and fertilized once a week.

Seed Treatment Before sowing, seeds were soaked in a methyl jasmonate (MeJA) solution of 0.8 mM for 24 h. For this, MeJA (Sigma Aldrich, São Paulo, Brazil) was dissolved in a small amount of ethanol (0.25 % in the final solution) and brought to the desired concentration with distilled water. As a control, another set of seeds was soaked in distilled water with the same amount of ethanol for 24 h. During the treatment, seeds were kept in the dark at room temperature.

Feeding Test 1stinstar T. absoluta larvae that had emerged within the previous 24 h were transferred in groups of two to plastic cups with a treated or untreated tomato leaflet. The leflets used were all the 3rd leaves from 4-5- weeks-old plants. The petioles were wrapped in cotton cloth to maintain turgidity. Every 3 days, leaflets were replaced to ensure fresh food for the larvae. Larvae were observed daily until they pupated. The duration of the larval period (in days) and the pupal weight (in mg) was recorded. The experiment was replicated 100 times (N=100) per treatment, all performed at once.

Collection and Analyses of VOCs VOCs from 1) control plants, 2) *T. absoluta*-infested plants, 3) MeJA-treated plants, 4) MeJA-treated+*T. absoluta*-infested plants were collected over a 24 h-period (at 26 ± 1 °C; L:D cycle 12:12 h). Before sampling, plants were removed from the pot, and the substrate was wrapped in aluminum foil to prevent collecting volatiles from the roots or the substrate. To induce volatiles, 15 newly emerged larvae were carefully transferred to the leaves of the

6-7-weeks-old plants and left for 24 h to allow larvae to settle and to enter the leaf parenchyma. On the sampling day (24 to 48 h after infestation), plants were individually placed in 1-L airtight glass chambers. The VOC collection and the analytical procedures were previously described by Martins and Zarbin (2013). Airflow (1 L/min) was introduced into the chamber, and VOCs were pushed and trapped onto 20 mg Havesep Q 80-100 mesh in a glass tube, and then eluted with 300 µl of double-distilled HPLC-grade hexane. Ten µl of tetradecane (50 ppm) were added as an internal standard (IS). Samples were concentrated to $100 \ \mu l$ by exposing them to a slow flow of synthetic air, and 1 μ l of the extract was injected in splitless mode and analyzed by GC/MS (Shimadzu QP 2010 Plus) with a RTX-5 column (30 m×0.25 mm i.d., 0.25 mm film thickness; Restek, Bellefonte, PA, USA). The column oven temperature was held at 40 °C for 1 min, increased to 250 °C at 7 °C min⁻¹, and held for 10 min. Helium was the carrier gas at a column head pressure of 170 kPa. Compounds were quantified based on IS peak area. Identification of individual compounds was achieved by comparing the calculated Kovats Indexes (KI) with the literature. In addition, for 2-hexen-1-ol, α -pinene, β -myrcene, α phellandrene, α -terpinene, β -phellandrene, nonanal, MeSA, decanal, β -caryophyllene, geranylacetone, and α -humulene, the mass spectra were compared with those from synthetic compounds, and for the others with mass spectra from the literature. A total of 7 replicates (N=7) per treatment were analyzed.

Behavioral Tests with Chrysoperla externa Dual-choice tests were conducted on a Y-tube olfactometer (Ø=2 cm; main arm=18 cm; smaller arms=13 cm) with a constant airflow of 0.5 L/min (each arm) previously humidified and passed through an active charcoal filter. Behavioral tests were conducted with the extracts from VOCs collected over 24 h as previously reported by Girling and Hassall (2008). As odor sources, one piece of filter paper (2×2 cm) with 10 µl of extract (VOC samples concentrated to 100 µl, as described above), were placed at the end of the smaller arms. To avoid dilution due to evaporation, a new odor sources was used for every observation. Third instar larvae of C. externa were starved for 24 h and then individually introduced at the distal end of the central Y-tube arm. The response was determined when the insect walked towards one of the odor sources and touched the filtered paper. After every 5 insects, the Y-tube olfactometer was rotated 180°. Larvae were observed for a maximum of 10 min. We chose this time period as a parameter based on preliminary tests. Previous behavioral tests with plant extracts on hexane as odor sources showed that the odor source remains attractive for that period of time (Seenivasagan et al. 2009). Insects that did not respond after this period of time were excluded from the analysis. We compared the response of larvae for the extracts between (i) undamaged

plants vs. *T. absoluta*-infested plants (N=40); (ii) undamaged MeJA-treated plants vs. *T. absoluta*-infested-MeJA-treated plants (N=30); (iii) *T. absoluta*-infested plants vs. *T. absoluta*-infested MeJA-treated plants (N=30). All three comparisons were tested each day in order to avoid results that were the effect of date on behavior.

Statistical Analyses Larval development and pupal weight of *T. absoluta* were analyzed using ANOVA. Data were checked for normality and were analyzed using Minitab 16.0 software (Minitab, State College, PA, USA). Individual VOCs emissions were transformed (Log X+2) and analyzed with a two-way ANOVA that included herbivory, MeJA, and their interaction as main factors. The preference of the predators in the olfactometer was analyzed by applying a two-sided binomial test. The VOC emissions and the preference of *C. externa* were analyzed using SPSS 14.0 for Windows (SPSS Inc., Chicago, IL, USA).

Results

Feeding Test Duration of the larval stage of *T. absoluta* was significantly longer for larvae fed on tomato plants treated with MeJA at the seed stage in comparison with the control (Fig.1a) (MeJA: F=229.61, df=1, P=0.001). On average, larvae feeding on leaves of MeJA-treated plants took 1.3 days longer to pupate in comparison with those fed control plants. Similarly, larvae reared on leaves from the treated plants showed lower pupal weights in comparison with those reared on the leaves from untreated plants (Fig.1b) (MeJA: F=341, df=1, P=0.001).

Emission of Volatile Organic Compounds by Tomato Plants Twenty eight compounds that include aliphatic and aromatic compounds as well as an array of terpenoids were identified in the headspace of tomato plants under the different treatments (Table 1). Herbivory alone significantly increased the emissions of α -pinene (P=0.016), 6-methyl 5-hepten-2one (P=0.011), β -myrcene (P=0.011), (E)- β -ocimene (P=0.019), isoterpinolene (P=0.021), TMTT (P=0.017), (Z)-3hexenyl butyrate (P < 0.001), decanal (P = 0.032), and hexyl salicylate (P=0.009). The emissions of several compounds from uninfested MeJA-treated plants decreased when compared to uninfested untreated plants. This was not the case for herbivore-infested plants, except for the emission of α cubebene (P=0.047), which was significantly decreased by the MeJA treatment in both uninfested and infested plants. In addition, the MeJA treatment significantly affected α -pinene (P=0.023 and) and β -myrcene (P=0.017). The results of the interactions suggest that herbivory affected the emissions of α -pinene (P=0.021), β -myrcene (P=0.012), γ -terpinene (P=

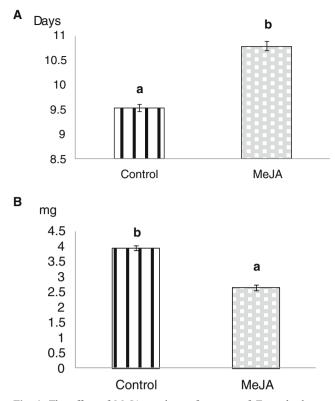


Fig. 1 The effect of MeJA on the performance of *Tuta absoluta*. **a** Length of development to pupation (days) and **b** Pupal weight (mg). Data are shown as means. *Error bars* represent the standard error of the mean, and the letters indicate statistical different means as determined byTukey's Test

0.010), and geranyl acetone (P=0.031) in the presence of the MeJA treatment (Table 2). While the MeJA treatment tended to reduce the emission from uninfested plants, the treatment tended to increase the emission of these compounds in the presence of herbivory.

Odor Preference of Chrysoperla externa Third instar larvae of *C. externa* oriented toward the odor of the extracts from *T. absoluta*-infested tomato plants in both untreated (P= 0.038) and MeJA-treated (P=0.001) plants over their respective controls. There was, however, no difference in the responses of the larvae to infested plant extracts that had been treated with MeJA or not (Fig. 2).

Discussion

The use of biological and chemical elicitors to induce resistance is considered a sound strategy for pest management programs (Stout et al. 2002). A good number of studies have addressed the effects of elicitor treatments at the seed stage to improve pathogen resistance, but only few have explored long lasting induced resistance against herbivores (Worrall et al.

2012). Seed treatments may offer advantages over foliar applications (Thaler 1999) or root drenches (Hodge et al. 2011) as they can be more cost-effective and reduce the negative consequences associated with pesticide use. Here, we addressed whether treating tomato seeds with MeJA affects the performance of T. absoluta, one of the major pests of tomato in South America, and whether the treatment can affect the preference of one of its main predators C. externa in this region. The results from the feeding test showed preliminary evidence that MeJA increases larval developmental time, which would reduce the number of generations per growing season. Because of the lengthened larval period, the larvae would also be exposed longer to predators and parasitoids, which would reduce pest populations (Price et al. 1980). In addition to increasing larval development time, the MeJA treatment reduced pupal weight. Pupal weight may be directly correlated with adult weight, which in turn is often correlated with fecundity and longevity of adult females as previously reported (Honěk 1993; Leather 1988), although this is not always the case (Fenemore 1977). Further studies are needed to understand how longer development and lower pupal weight caused by MeJA may affect T. absoluta population dynamics. Reduced performance of the herbivore may directly affect the performance of natural enemies due to decreased food quality. Treating plants of Vicia faba with a soil drench of β -amino butyric acid adversely affected the size of the endoparasitoid Aphidius ervi that developed in aphids fed treated plants (Hodge et al. 2011). Nevertheless, Hodge et al. (2011) suggest that predatory arthropods such as chrysopids may not be affected, as they may be able to compensate for lower food quality by increasing food consumption.

Among other factors, herbivory can increase plant volatile emissions (Dudareva et al. 2006). In tomato, significant increases of (E)- β -ocimene and TMTT occurred after herbivory by Tetranychus urticae (Acari), in addition to increases of linalool, (E)-nerolidol, and MeSA (Ament et al. 2004). In our study, the feeding by T. absoluta increased the emission of the terpenoids (E)- β -ocimene, isoterpinolene and TMTT and resulted in a slight, but not significant, increase of MeSA. We did not detect linalool or (E)-nerolidol in the headspace of tomato plants, which probably is due to the genetic characteristics of the studied cultivar. These two compounds have not been reported previously in the headspace volatiles from tomato cultivar. Santa Clara (Proffit et al. 2011). Our results, however, contrast with those of Thaler et al. (2002) who reported an increase in several compounds after herbivory by the polyphagous Lepidopteran species Spodoptera exigua. In addition to plant genotype and possibly different herbivorederived elicitors that lead to specific plant responses (De Moraes et al. 1998) that may result from differing feeding habits among herbivore species (Turlings and Wäckers 2004). Tuta absoluta is restricted to Solanaceae species and feeds on the parenchyma of the leaves, which results in leaf mines.

Table 1 Emissions of individual volatile organic compounds (Ng G Dw^{-1} 24 h^{-1}) by undamaged and *Tuta absoluta*-infested plants subjected to a methyl jasmonate treatment at the seed stage (Mean±S.E.M; N=7)

		Control				Herbivory			
	K.I.	MeJA-untreated		MeJA-treated		MeJA-untreated		MeJA-treated	
(<i>E</i>)-2-hexen-1-ol	n. c.	12.50	± 7.92	7.18 ^a		5.51 ^a		4.53 ^a	
(Z)-3-hexen-1-ol	n. c.	18.92	± 12.22	1.79 ^a		32.01	± 20.72	11.06	± 9.81
α- pinene	n. c.	120.02	± 25.41	25.17	± 10.51	121.52	± 27.52	134.32	± 31.29
6-methyl-5-hepten-2-one	951	32.02	± 9.28	9.67	± 4.35	35.31	± 5.83	34.18	± 7.74
β-myrcene	963	43.90	± 7.05	14.29	± 6.96	42.10	± 5.98	49.65	± 12.16
2-carene	1,003	2,290.15	± 493.48	500.86	± 98.45	1,979.13	± 497.13	1,939.53	± 336.02
α- phellandrene	1,009	144.82	± 48.97	32.08	± 9.41	174.76	± 56.55	190.45	± 53.47
α-terpinene	1,019	83.70	± 31.98	14.68	± 6.85	107.16	± 38.49	101.36	± 28.99
Limonene	1,032	768.42	± 253.39	219.92	± 52.58	871.43	± 207.82	894.33	± 234.26
β- phellelandrene	1,035	4,202.77	± 980.61	1,004.36	± 203.72	3,194.73	± 921.45	3,758.19	\pm 729.59
(E)- β -ocimene	1,048	9.08	± 4.49	3.80	± 2.47	36.12	± 13.38	25.58	± 10.56
γ-terpinene	1,062	13.41	± 3.75	3.23	± 1.57	13.57	± 5.78	19.32	± 2.69
Isoterpinolene	1,086	10.84	± 7.03	1.32 ^a		16.38	± 6.65	23.79	± 7.96
Nonanal	1,107	174.92	± 61.06	118.28	± 41.13	193.03	± 58.85	305.31	\pm 72.91
(Z)-3-hexenyl butyrate	1,184	1.36 ^a		0.52 ^a		38.11	± 14.50	26.40	± 11.72
MeSA	1,197	48.75	± 21.94	72.24	± 53.70	93.62	± 25.07	140.13	± 52.35
Decanal	1,206	352.76	\pm 78.46	160.38	± 58.48	570.53	±248.23	454.77	± 127.48
Δ -elemene	1,344	63.00	± 25.36	47.44	± 40.55	82.50	± 35.02	156.85	\pm 82.48
α-cubebene	1,385	8.12	± 2.77	0.24 ^a		11.93	± 5.67	4.79	± 1.87
Longifolene	1,397	14.80	± 6.95	6.87	± 4.50	12.84	± 6.66	14.70	± 5.47
β-caryophyllene	1,433	445.05	± 109.91	207.93	± 144.17	296.80	± 82.58	542.32	± 263.86
γ-elemene	1,438	7.46	± 4.82	4.41	± 2.89	6.76	± 3.32	7.31	± 3.90
Geranyl acetone	1,448	34.80	± 14.33	8.03	± 3.45	15.57	± 5.61	31.36	± 8.60
α-gurjunene	1,452	14.54	± 3.94	4.29	± 2.74	10.20	± 4.38	6.37	± 2.46
α-humulene	1,470	81.83	± 18.91	60.16	± 35.22	56.57	± 20.26	112.40	± 53.46
TMTT	1,575	1,102.53	\pm 394.14	615.73	± 323.18	2,277.27	± 486.01	1,897.53	± 655.25
Hexyl salicylate	1,685	17.26	± 11.17	5.32	± 3.46	19.15	± 2.44	15.30	± 3.07
(Z)-nerolidol	n. c.	4.83 ^a		6.29	±4.33	41.50	±35.03	13.90	± 10.60

S.E.M. Standard Error of the Mean, K.I. calculated Kovats Index, n.c. not calculated

^a Compound emitted by only one sample. S.E.M. was not calculated

Infestation period duration (Kant et al. 2004) and the low density of the infestation chosen (Girling et al. 2011), may also have affected the emission of induced compounds.

Surprisingly, the MeJA-treatment tended to decrease the emissions by undamaged tomato plants (Table 1). These results contrast with those observed in another study where tomato plants were treated at the seed stage with JA and showed increased emissions of TMTT compared to untreated plants as well as induction of methyl salicylate (Smart et al. 2013). In addition to the genotype, one major difference between the two studies in tomato is the concentration of elicitor used. Whereas we used 0.8 mM MeJA for inducing resistance of the future plants, Smart et al. (2013) used a much higher concentration (3 mM JA) of the elicitor. The MeJA concentration used in our study was selected based on previous studies

showing negative effects on the performance of *H. zea* and an assessment of physiological responses (germination percentage, seedling growth, days to ripeness, and average fruit weight/plant) of tomato cultivar Micro-Tom at Penn State University (Paudel *et al.* unpublished). Plants may respond differently to various elicitor concentrations. The response of parasitoids to JA-induced Brussels sprout plants, for instance, is dose-dependent (Bruinsma et al. 2009), which suggests that induced VOCs vary depending on the concentration of the elicitor applied. At the lower dose of 0.8 mM, plants may have allocated the exogenous MeJA to direct chemical or mechanical defenses, or to other metabolic processes that resulted in lower volatile emissions in the absence of herbivory.

To determine the effect on trichome density, we made a preliminary counting of the total numbers per mm² in 4 plants

 Table 2 P-values for main effects of the MeJA-treatment to seeds and herbivory (by *Tuta absoluta*) and their interactions on the emissions of individual compounds

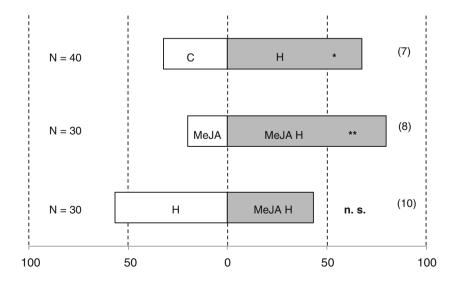
	MeJA	Herbivory	MeJA x Herbivory
(E)-2-hexen-1-ol	0.542	0.493	0.579
(Z)-3-hexen-1-ol	0.333	0.607	0.791
α-pinene	0.023	0.016	0.021
6-methyl-5-hepten-2-one	0.102	0.011	0.124
β-myrcene	0.017	0.011	0.012
2-carene	0.123	0.090	0.082
α -phellandrene	0.441	0.168	0.157
α-terpinene	0.463	0.122	0.203
Limonene	0.742	0.054	0.789
β- phellelandrene	0.171	0.133	0.051
(E) - β -ocimene	0.278	0.019	0.841
γ-terpinene	0.792	0.072	0.010
Isoterpinolene	0.898	0.021	0.276
Nonanal	0.330	0.152	0.597
(Z)-3-hexenyl butyrate	0.636	< 0.001	0.837
MeSA	0.926	0.061	0.710
Decanal	0.123	0.032	0.123
Δ -elemene	0.805	0.204	0.491
α-cubebene	0.047	0.300	0.226
Longifolene	0.708	0.617	0.180
β-caryophyllene	0.054	0.435	0.149
γ-elemene	0.741	0.688	0.613
Geranyl acetone	0.953	0.376	0.031
α -gurjunene	0.064	0.908	0.248
α-humulene	0.190	0.369	0.099
TMTT	0.377	0.017	0.584
Hexyl salicylate	0.279	0.009	0.880
(Z)-nerolidol	0.967	0.242	0.576

Fig. 2 Preference of Chrysoperla externa towards the odors of control (C) vs. herbivore-infested plants (H); MeJA treated plants (MeJA) vs. MeJA-treated herbivore-infested plants (MeJA H) and herbivore-infested vs. MeJA-treated herbivore-infested plants. Asterisks show the level of significance P<0.05 *; P<0.01 **, n.s. means "not significant". Numbers on the left show the total number of insects tested in each treatment (N). The number of non responsive insects is shown on the right side in brackets

from each treatment, but we did not find a clear difference between both MeJA treatments (control upper side 5.2 ± 1.1 trichomes/mm²; lower side 15.6±5.3 trichomes/mm²; MeJA upper side $7.4\pm2..3$ trichomes/mm²; lower side 16.8 ± 5.0 trichomes/mm² by Scanning Electron Microspy). This suggests that induction of trichomes did not occur in MeJA treated plants. Nevertheless, treating the seeds of tomato cultivar Micro-Tom with MeJA, has resulted in increased levels of polyphenol oxidase in the leaves, with higher levels at higher concentrations (tested up to 1 mM) (Paudel et al. unpublished), while low doses of MeJA may have triggered stomatal closure, which may restrict the release of volatile compounds (Akter et al. 2013). The fact that some volatile compounds decreased in MeJA-treated plants is interesting because it may result in tomato plants that are less apparent to herbivores that rely on volatile cues for host location (Bleeker et al. 2009; Proffit et al. 2011). This was not within the scope of our study, but deserves further investigation as it may also be exploited to manipulate herbivore behavior and reduce pest pressure.

Jasmonic acid is involved in the activation of both direct and indirect responses of plants to herbivores, in addition to its role in plant physiological functions. For MeJA-treated and untreated plants, similar emissions were observed for most of the compounds after herbivory (except for α -cubebene that decreased significantly in both uninfested and infested plants treated with MeJA). This suggests that MeJA treatment did not affect the storage pools and the induction of VOCs upon herbivory. Hypothetically, plant species whose response is based on quantitative differences between volatile blends emitted from herbivore-damaged and mechanically damaged plants have a high level of direct defenses (Dicke et al. 1998), which is indeed the case for tomato (Kennedy 2003).

The development of new techniques such as the induction of resistance should be assessed in a multitrophic context (Stenberg et al. 2010). The results from the bioassay suggest



that chemical differences recorded in the VOC emissions do not interfere with the foraging behavior of the predator *C. externa.* We did not find any enhanced attraction of the predator towards the extracts of MeJA-treated plants when compared to those from untreated plants after infesting them with *T. absoluta.* It is possible that the compounds that differed between treatments were relevant for the predator. Additionally, no effect on behavior was observed for a parasitoid wasp in olfactometer tests using plants treated (through soil drenches) with β -amino butyric acid (Hodge et al. 2011).

Smart et al. (2013) found that the treatment of tomato seeds with a higher dosage of JA resulted in the emissions of volatiles that attract predatory mites in the absence of herbivory. Based on the volatiles produced in our study (no induced volatile compounds in uninfested plants), we did not assess the response of *C. externa* to the extracts from control and MeJA-treated, uninfested plants. Increasing volatile emissions has been regarded as an approach to boost biological control in crop fields. Nevertheless, the inducibility of both direct and indirect defenses has been favored by evolution as it reduces physiological and ecological costs for the plant (Agrawal and Karban 1999), and therefore the induction of VOCs in the absence of the herbivore should be cautiously assessed.

For behavioral tests, extracts from VOCs collected over 24 h were used, and thus they were a mixture of VOCs collected over the photo- and scotophase. The biosynthesis and emission of several compounds is known to be lightdependent (Kesselmeier and Staudt 1999; Loughrin et al. 1994), and variations of day and night VOC emissions have particular ecological functions (De Moraes et al. 2001). With the exception of TMTT and methyl salicylate that were emitted in a light-dependent fashion, Faraq and Paré (2002) found that undamaged and herbivore-damaged tomato plants emit similar amounts of terpenoids and GLVs during the photo and scotophase. This suggests that release of terpenoids occurs from storage pools (e.g., compounds stored in trichomes). For the aim of our study, we do not consider that having collected VOCs over 24 h affected the results (all samples collected under the same lighting conditions). The fact that the predators were able to discriminate between the extracts of undamaged and damaged plants supports our results.

Our data provide evidence that a seed treatment with a low concentration of MeJA can be used to improve pest resistance, and that the chemical differences in VOCs do not alter the behavior of the predator *C. externa.* However, further studies are needed to understand whether the predator can be indirectly affected (*e.g.*, reduced development or fecundity due to reduced quality of the prey). The array of plant responses to chemical elicitors such as JA depends upon elicitor dosage, how it is applied to the plant, the plant genotype and age, and possibly, the intensity of herbivory (*e.g.*, herbivore density) that challenges the plant. Further research is needed to clarify

these responses in order to successfully integrate elicitor treatments into integrated pest management systems.

Acknowledgments The authors thank Liah A. D. Ferreira for treating all seeds used in the experiment, and for taking care of the plants before the experiments, Dr. Päivi Tiiva for statistical support, Dr. Brigida Souza for supplying *Chrsyoperla externa* eggs for starting the rearing, Flavia Krechemer for providing us with eggs and pupae of *Tuta absoluta*, and Samantha Husmann for support in the analysis of trichomes. Prof. Saskya van Nouhuys has kindly read the manuscript for improving the language. Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) – Proc. No. 401928/2012-8, Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) e Instituto Nacional de Ciências e Tecnologia (INCT) de Semioquímicos na Agricultura and USAID Cooperative Agreement No: EPP-A-00-0400016-00 are acknowledged for financial support. D.P-Z. is currently funded by the Program Science Without Borders.

References

- Agrawal AA, Karban R (1999) Why induced defenses may be favored over constitutive strategies in plants. In: Tollrian R, Harvell CD (eds) The ecology and evolution of inducible defenses. Princeton University Press, Princeton, pp 45–61
- Akter N, Okuma E, Sobahan MA, Uraji M, Munemasa S, Nakamura Y, Mori IC, Murata Y (2013) Negative regulation of methyl jasmonateinduced stomatal closure by glutathione in Arabidopsis. J Plant Growth Regul 32:208–215
- Ament K, Kant MR, Sabelis MW, Haring MA, Schuurink RC (2004) Jasmonic acid is a key regulator of spider mite-induced volatile terpenoid and methyl salicylate emission in tomato. Plant Physiol 135:2025–2037
- Bleeker PM, Diergaarde PJ, Ament K, Guerra J, Weidner M, Schütz S, De Both MTJ, Haring MA, Schuurink RC (2009) The role of specific tomato volatiles in tomato-whitefly interaction. Plant Physiol 151: 925–935
- Boughton AJ, Hoover K, Felton GW (2005) Methyl jasmonate application induces increased densities of glandular trichomes on tomato, *Lycopersicon esculentum*. J Chem Ecol 31:2211–2216
- Bruinsma M, Posthumus MA, Mumm R, Mueller MJ, van Loon JJA, Dicke M (2009) Jasmonic acid-induced volatiles of *Brassica* oleracea attract parasitoids: effects of time and dose, and comparison with induction by herbivores. J Exp Bot 60:2575–2587
- Buzi A, Chilosi G, De Sillo D, Magro P (2004) Induction of resistance in melon to *Didymella bryoniae* and *Sclerotinia sclerotiorum* by seed treatments with acibenzolar-S-methyl and methyl jasmonate but not with salicylic acid. J Phytopathol 152:34–42
- Conrath U, Beckers GJ, Flors V et al (2006) Priming: getting ready for battle. Mol Plant Microbe Interact 19:1062–1071
- De Moraes CM, Lewis WJ, Paré PW, Alborn HT, Tumlinson JH (1998) Herbivore-infested plants selectively attract parasitoids. Nature 393: 570–573
- De Moraes CM, Mescher MC, Tumlinson JH (2001) Caterpillar-induced nocturnal plant volatiles repel conspecie females. Nature 410:577– 580
- Dicke M, Takabayashi J, Posthumus MA, Schütte C, Krips OE (1998) Plant-phytoseiid interactions mediated by herbivore-induced plant volatiles: variation in production of cues and in responses of predatory mites. Exp Appl Acarol 22:311–333
- Dudareva N, Negre F, Nagegowda DA, Orlova I (2006) Plant volatiles: recent advances and future perspectives. Crit Rev Plant Sci 25:417– 440

- El-Wakeil N, Gaafar N, Sallam A, Volkmar C (2013) Side effects of insecticides on natural enemies and possibility of their integration in plant protection strategies. In: Trdan S (ed). Insecticides— Development of Safer and More Effective Technologies, InTech, pp. 3–56.
- Embrapa (2014) Cultivo de Tomate para Industrialização http:// sistemasdeproducao.cnptia.embrapa.br/FontesHTML/Tomate/ TomateIndustrial_2ed/index.htm Accessed 9 June 2014.
- Faraq MA, Paré PW (2002) C6-Green leaf volatiles trigger local and systemic VOCemissions in tomato. Phytochemistry 61:545–554
- Farmer EE, Ryan CA (1992) Octadecanoid precursors of jasmonic acid activate the synthesis of wound-inducible proteinase inhibitors. Plant Cell 4:129–134
- Farooq M, Aziz T, Basra SMA, Cheema MA, Rehman H (2008) Chilling tolerance in hybrid maize induced by seed priming with salicylic acid. J Agron Crop Sci 194:161–168
- Felton GW, Donato K, Del Vecchio RJ, Duffey SS (1989) Activation of plant foliar oxidases by insect feeding reduces nutritive quality of foliage for noctuid herbivores. J Chem Ecol 15:2667–2694
- Fenemore P (1977) Oviposition of potato tuber moth, *Phthorimaea operculella* Zell. (Lepidoptera: Gelechiidae); fecundity in relation to mated state, age, and pupal weight. N Z J Zool 4:187–191
- Ghoneim K (2014) Predatory insects and arachnids as potential biological control agents against the invasive tomato leafminer, Tuta absolutaMeyrick(Lepidoptera: Gelechiidae), in perspective and prospective. J Entomol Zool Stud 2:52–71
- Girling RD, Hassall M (2008) Behavioural responses of the seven-spot ladybird *Coccinella septempunctata* to plant headspace chemicals collected from four crop *Brassicas* and *Arabidopsis thaliana*, infested with *Myzus persicae*. Agric For Entomol 10:297–306
- Girling RD, Stewart-Jones A, Dherbecourt J, Staley JT, Wright DJ, Poppy GM (2011) Parasitoids select plants more heavily infested with their caterpillar hosts: a new approach to aid interpretation of plant headspace volatiles. Proc R Soc B 278:2646–2653
- Hodge S, Galster AM, Ward JL, Beale MH, Powell G (2011) The effects of a plant defence priming compound, β-aminobutyric acid, on multitrophic interactions with an insect herbivore and a hymenopterous parasitoid. BioControl 56:699–711
- Honěk A (1993) Intraspecific variation in body size and fecundity in insects: a general relationship. Oikos 483–492
- Tuta absoluta Information Network (2014) http://www.tutaabsoluta.com/ tuta-absoluta. Accessed 29 January 2014.
- Kant MR, Ament K, Sabelis MW, Haring MA, Schuurink RC (2004) Differential timing of spider mite-induced direct and indirect defenses in tomato plants. Plant Physiol 135:483–495
- Karban R (2011) The ecology and evolution of induced resistance against herbivores. Funct Ecol 25:339–347
- Karban R, Baldwin IT (1997) Induced responses to herbivory. University of Chicago Press, Chicago
- Kennedy GG (2003) Tomato, pests, parasitoids and predators: Tritrophic interactions involving the genus *Lycopersicon*. Annu Rev Entomol 48:51–72
- Kesselmeier J, Staudt M (1999) Biogenic volatile organic compounds (VOC):an overview on emission, physiology and ecology. J Atmos Chem 33:23–88
- Kessler A, Baldwin IT (2002) Plant responses to insect herbivory: the emerging molecular analysis. Annu Rev Plant Biol 53:299–328
- Krantev A, Yordanova R, Janda T, Szalai G, Popova L (2008) Treatment with salicylic acid decreases the effect of cadmium on photosynthesis in maize plants. J Plant Physiol 165:920–931
- Leather SR (1988) Size, reproductive potential and fecundity in insects: things aren't as simple as they seem. Oikos 51:386–389
- Li L, van Staden J, Jäger AK (1998) Effects of plant growth regulators on the antioxidant system in seedlings of two maize cultivars subjected to water stress. Plant Growth Regul 25:81–87

- Lietti MM, Botto E, Alzogaray RA (2005) Insecticide resistance in Argentine populations of *Tuta absoluta* (Meyrick)(Lepidoptera: Gelechiidae). Neotrop Entomol 34:113–119
- Loughrin JH, Manukian A, Heath RR, Turlings TCJ, Tumlinson JH (1994) Diurnal cycle of emission of induced volatile terpenoids by herbivore-injured cotton plants. Proc Natl Acad Sci U S A 91: 11836–11840
- Martins CB, Zarbin PHG (2013) Volatile organic compounds of conspecific-damaged *Eucalyptus benthamii* influence responses of mated females of *Thaumastocoris peregrinus*. J Chem Ecol 39:602– 611
- Mithöfer A, Boland W (2012) Plant defense against herbivores: chemical aspects. Annu Rev Plant Biol 63:431–450
- Pankaj, Muttucumaru N, Powers SJ, Gaur HS, Kurup S, Curtis RHC (2013) Differential defence response due to jasmonate seed treatment in cowpea and tomato against root-knot and potato cyst nematodes. Nematology 15:15–21
- Pinto-Zevallos DM, Zarbin PHG (2013) A química na agricultura: perspectivas para o desenvolvimento de tecnologias sustentáveis. Quim Nova 36:1509–1513
- Pinto-Zevallos DM, Martins CB, Pellegrino AC, Zarbin PHG (2013) Compostos orgânicos voláteis na defesa induzida das plantas contra insetos herbívoros. Quim Nova 36:1395–1405
- Price PW, Bouton CE, Gross P, McPheron BA, Thompson JN, Weis AE (1980) Interactions among three trophic levels: influence of plants on interactions between insect herbivores and natural enemies. Annu Rev Ecol Syst 11:41–65
- Proffit M, Birgersson G, Bengtsson M, Reis R Jr, Witzgall P, Lima E (2011) Attraction and oviposition of *Tuta absoluta* females in response to tomato leaf volatiles. J Chem Ecol 37:565–574
- Seenivasagan T, Chander S, Navarajan Paul AV (2009) Orientation and behavioural responses of *Cotesia plutellae* Kurdjumov (Hymenoptera: Braconidae) to cruciferous host plants and host larval body extracts. J Biol Control 23:365–373
- Shakirova FM, Sakhabutdinova AR, Bezrukova MV, Fatkhutdinova RA, Fatkhutdinova DR (2003) Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. Plant Sci 164:317– 322
- Silva GA, Picanço MC, Bacci L, Crespo AL, Rosado JF, Guedes RN (2011) Control failure likelihood and spatial dependence of insecticide resistance in the tomato pinworm, Tuta absoluta. Pest Manag Sci 67:913–920
- Siqueira H, Guedes R, Picanço MC (2000) Cartap resistance and synergism in populations of *Tuta absoluta* (Lep., Gelechiidae). J Appl Entomol 124:233–238
- Smart LE, Martin JL, Limpalaër M, Bruce TJ, Pickett JA (2013) Responses of herbivore and predatory mites to tomato plants exposed to jasmonic acid seed treatment. J Chem Ecol 39:1297–1300
- Smith JL, De Moraes CM, Mescher MC (2009) Jasmonate- and salicylate-mediated plant defense responses to insect herbivores, pathogens and parasitic plants. Pest Manag Sci 65: 497–503
- Stenberg JA, Lehrman A, Björkman C (2010) Uncoupling direct and indirect plant defences: novel opportunities for improving crop security in willow plantations. Agric Ecosyst Environ 139:528–533
- Stout MJ, Zehnder GW, Bauer ME (2002) Potential for the use of elicitors of plant resistance in arthropod management programs. Arch Insect Biochem Physiol 51:222–235
- Thaler JS (1999) Jasmonate-inducible plant defences cause increased parasitism of herbivores. Nature 399:686–688
- Thaler JS, Farag MA, Paré PW, Dicke M (2002) Jasmonate-deficient plants have reduced direct and indirect defences against herbivores. Ecol Lett 5:764–774
- Traw BM, Dawson TE (2002) Differential induction of trichomes by three herbivores of black mustard. Oecologia 131:526–532

- Turlings TCJ, Wäckers F (2004) Recruitment of predators and parasitoids by herbivore-injured plants. In: Cardé RT, Millar JG (eds) Advances in insect chemical ecology. Cambridge University Press, Cambridge, pp 21–75
- van Dam NM, Witjes L, Svatoš A (2004) Interactions between aboveground and belowground induction of glucosinolates in two wild *Brassica* species. New Phytol 161:801–810
- van Poecke RMP, Dicke M (2002) Induced parasitoid attraction by Arabidopsis thaliana: involvement of the octadecanoid and the salicylic acid pathway. J Exp Bot 53:1793–1799
- Villas Bôas GL, Castelo Branco M, Medeiros MA (2009) Manejo integrado da traça-do-tomateiro (Tutaabsoluta) em sistema de produção integrada de tomate indústria (PITI). 16 p. Brasília: Embrapa Hortaliças (Circular Técnica 73)
- Worrall D, Holroyd GH, Moore JP, Glowacz M, Croft P, Taylor JE, Paul ND, Roberts MR (2012) Treating seeds with activators of plant defence generates long-lasting priming of resistance to pests and pathogens. New Phytol 193:770–778
- Wu J, Baldwin IT (2009) Herbivory-induced signalling in plants: perception and action. Plant Cell Environ 32:1161–1174

Copyright of Journal of Chemical Ecology is the property of Springer Science & Business Media B.V. and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.